Journal of Pharmaceutical Research International

33(41A): 26-40, 2021; Article no.JPRI.72650 ISSN: 2456-9119 (Past name: British Journal of Pharmaceutical Research, Past ISSN: 2231-2919, NLM ID: 101631759)

Bone, it's Architecture, Markers of Resorption and Their Role in Osteoporosis in India: A Detailed Review

Tirthal Rai¹ , Janice DSa2* and Premjit Sujir3

1 Department of Biochemistry, Nitte (Deemed to be University), K.S Hegde Medical Academy (KSHEMA), Mangalore, 575018, India. ² Department of Biochemistry, A.J Institute of Medical Sciences & Research Centre, Mangalore, 575004, India. 3 Department of Orthopaedics, Kasturba Medical College, Manipal Academy of Higher Education (MAHE), Mangalore, Karnataka, 575001, India.

Authors' contributions

This review article was written in collaboration with all three authors. Author TR designed the study and wrote the study protocol. Author JD drafted the manuscript. Author PS managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JPRI/2021/v33i41A32299 *Editor(s):* (1) Dr. Sawadogo Wamtinga Richard, Ministry of Higher Education, Scientific Research and Innovation, Burkina Faso. *Reviewers:* (1) Md. Munzur Rahman, Rajshahi Medical College, Bangladesh. (2) Satyam Bhodaji, MGM School of Physiotherapy, India. (3) Evelyn Sharon S, Dr. MGR Educational and Research Institute, India. (4) Elsayed Morsi, Menoufia University, Egypt. Complete Peer review History: https://www.sdiarticle4.com/review-history/72650

Review Article

Received 08 June 2021 Accepted 12 August 2021 Published 14 August 2021

ABSTRACT

Bone is a dynamic tissue that undergoes constant remodeling throughout the life span. Bone turnover is an equilibrium between the rates of bone formation and resorption. Assay of bone turnover markers (BTMs) is very important as they provide an insight in to the dynamics of bone turnover in many metabolic bone disorders. An increase in bone turnover seen with aging and pathological states such as osteoporosis leads to deterioration of bone microarchitecture and thus contributes to an increase in the risk of fracture independent of low bone mineral density (BMD). These microarchitectural alterations affecting the bone quality can be assessed by BTMs and thus may serve as a complementary tool to BMD in the assessment of fracture risk. Osteoporosis is a major health problem in India with 230 million people getting affecting. Women get more crippled as

**Corresponding author: E-mail: drjanicedsa@ajims.edu.in;*

they show an additional accelerated phase of bone loss, which occurs 10 years earlier in India. Bone health in Indian women is more appalling with deficient nutritional status as compared to their counterparts. Biochemical assays are non-invasive, not site specific, reflect the turnover of entire skeleton, can detect early changes in the bone turnover. The present review was aimed to discuss the normal architecture of bone, markers of it's turnover and their role in osteoporosis in India.

Keywords: Bone turnover markers; BTM; osteoporosis; India.

1. INTRODUCTION

Osteoporosis is a progressive metabolic or skeletal disorder, leading to a consequent increase in fracture risk due to deterioration in bone mass and imperfection in its microarchitecture. Osteoporosis is defined as bone mineral density that is less than 2.5SD below the reference range in young adults of the same race and sex (t score of $- 2.5$) [1].

Bone grows in size during the first two decades of life as anticipated, with an accelerated growth during adolescence precisely related to estrogen in females and testosterone in males. In the third decade individuals attain maximum bone mass $[1]$.

Approximately 3 % of cortical bone is replaced each year and 25% of trabecular bone is resorbed and replaced each year. (Fig 1) Women of all ethnic groups show an additional accelerated phase of bone loss, which occurs for

about 10 years after the cessation of ovarian function. Total bone loss in osteoporosis may exceed 30 to 40%. In women there is a 15 to 18% lifetime risk of hip fracture after the age of 50 years, versus 6% risk in men [3]. 230 million people in India are over the age of 50 and vulnerable to fragility fractures [4].

Normal bone density is defined as -1 standard deviation or greater than the mean at $30 - 40$ years (peak bone mass). Bone density between - 1SD and -2.5 SD of peak bone mass is osteopenia and less than 2.5 SD of peak bone mass is osteoporosis. For the diagnosis of osteoporosis, it has been recommended by the international osteoporosis foundation (IOF) to measure Bone mineral density (BMD) by dual X ray absorptiometry at the hip and lumbar region, and by this definition general prevalence of osteoporosis rises from 5% in women at the age of 50 years to 50% at the age of 85years and in men the comparable figures are 2.4% and 20% [5].

Fig. 1. Normal trabecular bone (top) / Osteoporotic trabecular bone (bottom) [2]

Osteoporosis is a silent killer, affecting more than one million women over the age of 50. Approximately 75% of the population faces the risk of fractures. Every eighth man and every third females in India suffers from osteoporotic fractures ten years earlier as compared to other developed countries, making India one of the
largest affected countries globally. Life largest affected countries globally. Life expectancy in India is up to 67 years which is expected to increase to 71 years in 2025. With increase in longevity there will be a 34% escalation in elderly population making them more susceptible to osteoporotic fractures. Osteoblasts, osteoclasts and osteocytes play a crucial role in bone remodeling [6]. This is a highly complex process comprising of coordinated action of these bone cells. In osteoporosis, the coordination is hampered and the basic multicellular unit (BMU) is influenced. This in turn is measured by bone turnover markers (BTM). Recent studies have suggested BTMs to outshine as markers for diagnosis of osteoporosis, these biochemical reflect the turnover of entire skeleton and can detect early changes in bone turnover [7]. The present nonsystematic review was aimed to discuss the normal architecture of bone, markers of it's turnover and their role on osteoporosis in Indians.

2. BONE ARCHITECTURE

2.1 Histology of Bone

Normal bone is lamella. It can be cortical or cancellous. Cortical bone makes up 80% of the skeleton and is composed of tightly packed osteons that are connected by Haversian canals. These canals contain arterioles, venules, capillaries and nerves. Interstitial lamellae lie between the osteons. Cement lines define the outer border of the osteon, cortical bone is characterized by slow turnover rate and cancellous bone is less dense and undergoes more remodeling according to the lines of stress. Periosteum is a thick connective tissue comprising of blood vessels, nerves and osteogenic cells and they pack the cortical bone, whereas endosteum is a thin layer of connective tissues covering the cavities [8]. It has a higher turnover rate and is more elastic than cortical bone. During osteoporosis both cortical and trabecular bone are affected making the bones brittle and more prone to fractures. The trabecular bone is composed of plates and rods providing strength. As age progresses plates become more rod like and the connectivity

between them is lost, which lowers the stiffness and bone strength. This arrangement is evident in femoral neck. Inferior cortex provides resistance to compressive arcade and superior cortex resists tensile load. This conjunction provides flexibility and strength [9]. Failure of this interconnectivity is the reason for femoral neck fractures [10]. Cortical porosity also defines bone strength and is independent of cortical size. Large macropores are created by increased osteoclastic activity by expanding the Haversian canal. This will lead to incapability in bearing loads by thinning the cortical tissue. This mechanism with increased spaces and macropores is commonly seen in old age. The proximity in these macropores minimizes the impact on bone strength [11].

2.2 Cellular Biology [Fig 2]

Osteoblasts: Osteoblasts are derived from undifferentiated mesenchymal cells. These cells form bone. These cells have more endoplasmic reticulum, Golgi apparatus, and mitochondria. More differentiated, metabolically active cells, line bone surfaces and less active cells in the resting regions maintain the ionic milieu of bone. Disruption of the lining layer activates osteoblasts. This osteoprogenitor lineage is activated by gene expression, timely programmed steps such as formation of bone morphogenetic protein (BMPs) [12]. Gene expression of Runt related transcription factor 2(Run X2) is the most crucial gene of osteoblast differentiation. RunX2 devoid mice have demonstrated to down regulate osteoblast related gene ColIA1, Cbf alpha and Cbf beta. Cbf beta is the most recent gene noticed in bone marrow and plays a vital role in osteoblast differentiation, followed by proliferation phase where, osteoblasts show high alkaline phosphatase activity, produce type I collagen, respond to parathyroid hormone and produce osteocalcin [13][14]. Osteoblasts have receptor effector interactions for parathyroid hormone, vitamin D3, glucocorticoids, prostaglandins and estrogen.

Osteocytes: Make up 90% of the cells in the mature skeleton and serve to maintain bone. These cells represent former osteoblasts that have been trapped within newly formed matrix. The embedded osteocytes differ in their morphology depending on the bone type. trabecular bone osteocytes are round whereas osteocytes from cortical bone are comparatively flatter [16]. They have an important role in controlling the extracellular concentration of calcium and phosphorous. They are directly stimulated by calcitonin and inhibited by parathyroid hormone. Once the osteocyte gets trapped within the mineralized bone matrix, the osteoblast markers such as osteocalcin and alkaline phosphatase are downregulated while expression of osteocyte markers such as dentine matrix protein1 (DMP1) and sclerostin are increased [17]. Osteocytes detect mechanical pressures and loads due to the interconnected network, thereby helping the bone to adapt to daily mechanical forces. The regulation of osteoblasts and osteoclasts mechanism depicts osteocytes as orchestrators of bone remodelling [18].

Osteoclasts: These multinucleated, irregularly shaped giant cells originate from hematopoietic tissues involved in resorption of bone. Multiple factors promote activation of gene expression in osteoclasts, factors such as macrophage colonystimulating factor (M-CSF) and RANK ligand which is secreted by osteoblasts, osteocytes, and stromal cells play a crucial role in osteoclastic activity [19]. Osteoprotegerin (OPG), which is produced by osteoblasts, stromal cells, gingival and periodontal fibroblasts bind to RANKL, preventing osteoclastogenesis by inhibiting the interaction of RANK/RANKL. These interactions further promote gene expression of DC-STAMP, which plays a critical role in fusion of osteoclastic cells [20]. Bone resorption occurs in depressions known as Howships lacunae by osteoclasts that synthesize tartrate resistant acid phosphate and cathepsin K. Osteoclasts produce hydrogen ions via carbonic anhydrase to lower *Rai et al.; JPRI, 33(41A): 26-40, 2021; Article no.JPRI.72650*

the pH, which increases the solubility or Hydroxyapatite crystals, and organic matrix is removed by proteolytic digestion. Osteoclasts also have specific receptors for calcitonin to allow them to directly regulate bone resorption [21].

3. OSTEOPROGENITOR CELLS

These local mesenchymal cells line the haversian canals and the periosteum, awaiting the stimulus to differentiate into osteoblasts. In bones that have stopped displaying bone remodeling or when matured are spindle shaped and are "inactive osteoblasts". They are exhibited frequently and in enormous numbers in developing bones. These cells also reside in perichondrium and escalate the bone morphogenetic proteins that independently help in formation of bone matrix. Recent studies have
confirmed that osteoprogenitor cells that confirmed that osteoprogenitor originate from hypothalamus increase osteogenesis and inhibitors of proline rich tyrosine kinase 2 regulates early osteoprogenitors and can be used in the treatment of osteoporosis [22].

Cells: These are narrow, flattened cells that form an envelope around the bone. Functions of bone lining cell is not well understood. However, they prevent the interaction of osteoclasts and bone matrix during bone formation and produce OPG and RANKL that participate in osteoclast differentiation. Lining cell equally play a pivotal role in bone remodeling as other bone cells [23- 24].

Fig. 2. Schematic diagram of bone turnover [15]

4. BONE MATRIX

Organic components: Organic components make up 40% of the dry weight of bone. They include Collagen, Proteoglycans, Non collagenous matrix proteins and Growth factors and Cytokines. Collagen is responsible for the tensile strength of the bone and makes up 90% of the organic matrix of bone. It is composed primarily of type I collagen. Collagen structure consists of a triple helix of two α 1 and one α 2 chains. Small leucine rich proteoglycans like decorin, biglycan, lumican, osteoaderin, and seric proteins are partially responsible for the compressive strength of the bone [25]. There are a number of non-collagenous matrix proteins that promote mineralization and bone formation. They include Osteocalcin which is produced by osteoblasts, is directly related to the regulation of bone density. The synthesis of osteocalcin is inhibited by PTH and stimulated by Vit D3. Osteocalcin levels can be measured in the serum or urine as a marker of bone turnover. Osteonectin is secreted by platelets and osteoblasts. It is postulated to play a role in the regulation of calcium metabolism. Osteopontin is a cell binding protein and Growth factors and cytokines: Transforming growth factor β, Insulin like growth factor, Interleukins 1 & 6 and bone morphogenic proteins [26].

Inorganic components: Make up 60% of the dry weight of bone. Calcium hydroxyapatite makes up most of the inorganic matrix of bone and is responsible for the mineralization of the matrix. Osteocalcium phosphate {BRUSHITE}, makes up the remaining inorganic matrix. It also has significant amount of bicarbonate, sodium, potassium, citrate, magnesium, carbonate, fluorite, zinc, barium, and strontium are also present. The conjunction of these inorganic components with collagen and non-collagenous matrix forms a hydroxyapatite, providing strength and mechanical support to the bone matrix. These complexed frame work executes an important role in bone homeostasis and prediction of bone fractures. Age, nutrition, disease, and osteoporotic treatments have exhibited diverse effects on bone matrix proteins that cause detrimental effects on bone structure. Bone matrix contributes to bone quality [27].

5. BONE CELLS AND BONE MATRIX

Bone matrix acts as a ground substance over which the bone cells anchor. They regulate the activity of bone cells through adhesion molecules *Rai et al.; JPRI, 33(41A): 26-40, 2021; Article no.JPRI.72650*

like integrins [28]. These cause osteoblasts to interact with bone matrix by binding to proteins like fibronectin, Osteopontin, collagen etc. integrins of osteoblasts are α 1 β 1, α 2 β 1, and α 5 β 1 and they play a pivotal role on organisation of osteoblasts on bone matrix during osteoid synthesis. The interaction between osteoclasts and bone matrix is very essential for exhibiting its role. Thus, during osteoclastic activity $\alpha\gamma\beta$ and α 2 β 1 integrins bind to bone-enriched proteins, such as bone sialoprotein and Osteopontin. Despite these bindings, cadherins that provide constant contact between osteoclast and growth factors required for osteoclast differentiation fail to express. β 3 and β 1 integrins are involved in interaction of osteocytes to bone matrix. These are crucial as they induce signals and amplify them on detecting bone tissue deformity [29].

6. OSTEOPOROSIS IN INDIA

6.1 Risk Factors for Osteoporosis

The data obtained is mainly done by studies scattered in different parts of India.

In 2015, 230 million Indians are expected to be over 50 years with 20% sustaining osteoporotic fractures. Women of all ethnic groups are more prone to low bone mass after menopause. Estrogen hormone is required for bone formation and growth. Age of menopause in Indian women is ten years earlier as compared to Caucasian women [30-31]. Genetic factors also correlate strongly with bone mass. Indian women have shown lower BMD than other ethnic counterparts. It is also thought that vitamin D receptor polymorphism is responsible for varied results [32].

Even though India has ample sunlight, vitamin D deficiency is observed in Indians regardless of their gender. Diet, increase melanin production and attire in India may hinder the formation of Vitamin D. As the importance of vitamin D in bone growth is discussed earlier, osteoporosis is strongly influenced by this hormone. Thus, indirectly leading to demineralization and tampering the bone matrix. Indians also lack the daily recommended intake of calcium. The Indian diet hinders calcium absorption as it is rich in phytates. Thus, hindering the formation of bone matrix [33].

Sedentary life style in elders decreases exposure to sunlight and lack of exercise decreases muscle activity and acts as stressor to escalate bone formation, thus improving bone quality [34]. Alcoholism and cigarette smoking can also cause detrimental effect on bone health. Smoking increases the lifetime risk of developing fragility fracture is 13% and 32% in women and men respectively [35]. However, significant relation of this parameter to osteoporosis in India is questionable, as the frequency of smokers in India are comparatively less. Long term treatment on glucocorticoids is a major contributor to osteoporosis in all ethnic groups [36]. Body mass index is a crucial risk factor for osteoporosis. However, it is controversial and contradicting. Obese women are said to have a protective effect on osteoporosis. Adipose tissue and BMD are positively correlated in many studies. However poor nutritional status can have detrimental effect on bone mass [37].

7. CLINCAL FEATURES OF OSTEOPOROSIS

It is noticed only after the first fracture. Most common fractures associated with osteoporosis in India are.

Vertebral fracture: Majority of vertebral fractures result from compressive loading associated with activities such as lifting, changing position, coughing sneezing or a simple fall from standing height. However only one third of new vertebral fracture relates to fall. The earliest and the major symptom of vertebral compression fracture is acute back pain and the most routine activities could be restricted. 17.1% of vertebral fractures in 415 female subjects in India were reported at the Delhi vertebral osteoporosis study [38].

Hip fracture: The incidence of hip fractures increases exponentially with age and is expected to escalate to 6.3 million in 2050. Over the age of 50, hip fractures were estimated to be 159/1000 women. Indian studies revealed that men had higher chance of sustaining hip fractures as compared to the preponderance of these fractures in females [39]. Hip fractures mainly involve the femoral neck and intertrochanter fractures in the elderly osteoporotic population, which are caused by low trauma. These fracture cause excruciating pain, loss of mobility and excess mortality. Nearly all are hospitalized and most undergo surgery. It is estimated that medical expenditures for hip fractures are exponentially high. However, non-operative treatment can lead to disastrous complications and operative treatment like arthroplasty and

internal fixation are prone to failures that can cause irreparable damage.

8. BONE TURNOVER MARKERS

Measurement of Bone mineral density is at present the method of choice in the evaluation of fracture risk. However, they are influenced by multiple factors. Rural India has limited set ups for DEXA in government hospitals therefore suggesting scanty access to these machines [43]. Increasing evidence suggest that the risk of osteoporosis and fracture is associated with increase bone turnover as measured by markers of resorption and formation, independently. At present, the markers reflecting bone formation with acceptable specificity and sensitivity are osteocalcin, bone specific alkaline phosphatase and procollagen type 1 N-terminal pro-peptide. For evaluating bone resorption, pyridinoline, deoxy pyridinoline, and N and C terminal type 1 collagen telopeptide, urinary hydroxyproline, urinary excretion of the pyridinium cross-links (pyridinoline and deoxypyridinoline), and plasma tartrate-resistant acid phosphatase activity [44].

8.1 Biochemical Markers of Bone Formation

Serum alkaline phosphatase: Serum alkaline phosphatase activity is the most commonly used marker of bone formation, but it lacks sensitivity and specificity, especially in patients with osteoporosis. In an attempt to improve the specificity and the sensitivity of serum alkaline phosphatase measurement, techniques have been developed to differentiate the bone and the liver isoenzymes, which differ only by posttranslational modifications since they are coded by a single gene. These techniques rely on the use of differentially effective activators and inhibitors (heat, phenylalanine, and urea), separation by electrophoresis, and separation by specific antibodies. In general, these assays have slightly enhanced the sensitivity of this marker, but most of them are indirect or technically cumbersome or both. A real improvement should be obtained by using a monoclonal antibody recognizing the bone but not the liver and kidney isoenzyme. Serum total ALP has revealed interaction with spinal bone matrix. Recent study depict ALP as a marker for osteoporosis in men. Indian studies also revealed the same with significant higher levels in osteoporotic population [45].

Fig. 3. X rays showing osteoporotic fracture of proximal femoral neck (1a), middle femoral femoral Fig. 3. X rays showing osteoporotic fracture of proximal femoral neck (1a), middle femoral
neck (1b), basilar femoral neck (1c) and inter and subtrochanteric femoral regions. (1d) [40] *Distal forearm fracture: among the fractures sustained in osteoporosis, this fracture seems to be economically and emotionally less effective. However, it is speculated that these fractures can lead to fragility fractures in future, which is economically draining. Most distal forearm fractures occur in women. The age adjusted female to male ratio being 4: 1. there is a rapid increase in the incidence after menopause which plateaus at about 65* and emotionally less effective. However, it is speculated that these fractures can lead to fragility fractures in
ure, which is economically draining. Most distal forearm fractures occur in women. The age adjusted female
m **orearm fracture:** among the fractures
motionally less effective. However, it is
vhich is economically draining. Most dis
ratio being 4: 1. there is a rapid increas

Fig. 4. X ray showing distal radius fracture [42] ray

Serum osteocalcin (OC): Osteocalcin, also called bone gla protein (BGP), is a small, non collagenous protein that is specific for bone tissue and dentin, but its precise function remains unknown. Osteocalcin is synthesized predominantly by the osteoblasts and incorporated into the extracellular matrix (ECM) of bone, They are gene mediated and the gene encoding for OC is the BGLAP gene. The physiological role of osteocalcin is glucose and calcium homeostasis, bone mineralisation and bone remodelling. Osteocalcin originally is

Frum osteocalcin (OC): Osteocalcin, also synthesized as a prohormone with 95 amino
lled bone gla protein (BGP), is a small, non acids and then cleaved to form a mature
llagenous protein that is specific for bone polypept acids and then cleaved to form a mature polypeptide consisting of 49 amino acids in humans. Comprising of three gamma carboxy glutamic acid residues at positions 17, 21 and 24. This binds the osteocalcin to calcium and exhibits a helical structure with compact calcium synthesized as a prohormone with 95 amino
acids and then cleaved to form a mature
polypeptide consisting of 49 amino acids in
humans. Comprising of three gamma carboxy
glutamic acid residues at positions 17, 21 and
24. Thi molecule in the extracellular matrix of the bone [46]. Osteocalcin is activated by Vitamin K, a co factor for gamma carboxylation. OC circulates in the blood stream in two forms carboxylated and acellular matrix of the bone
activated by Vitamin K, a co
rboxylation. OC circulates in
two forms carboxylated and

uncarboxylated OC but a fraction of newly synthesized carboxylated OC is released into the circulation, where it is predominantly measured by radioimmunoassay. Uncarboxylated OC requires sophisticated techniques. Depending on the epitopes recognized by the antibody, monoclonal antibodies are used against OC fragments such as N-mid and N terminal OC fragments and in addition intact OC fragments of osteocalcin. Some of these fragments may be released during bone resorption in an acidic condition during high turnover state, but some of them are unrelated to resorption and their significance is not known. In most cases, however, serum osteocalcin is a valid marker of bone turnover when resorption and formation are coupled and is a specific marker of bone formation whenever formation and resorption are uncoupled. Osteocalcin is dependent on post – translational carboxylation for its hydroxyapatite affinity, a process that has been shown to decrease with age. High levels of under carboxylated osteocalcin are seen in hip fracture patients. OC is cost effective and can be used in screening and management of low bone mass in Indian population, as they are >85% sensitive, accurate and precise in diagnosing [47].

Procollagen type I propeptides (PINP): They are derived from collagen type I, in which they form amino- (N-) and carboxy- (C-) terminal extension peptides. Since both the carboxy- and the amino-terminal propeptides of type I collagen are generated in a stoichiometric fashion, the propeptides are considered quantitative measures of newly formed type I collagen. Both propeptides may be measured by specific, polyclonal-based immunoassay. PINP originates from fibroblasts and proliferating osteoblasts, also derived from skin, tendon, dentin, and cartilage. PINP is a better bone formation marker than its counterpart and is proposed as a novel bone formation marker by International osteoporosis foundation in account of its response to treatment, precision and reliability as evidenced by low variability and longer stability at room temperature. A study correlating PINP levels with age and sex matched Indians suggested the same with no gender difference and no variations with vitamin D deficiency [48].

8.2 Bone Resorption Markers

Most biochemical markers of bone resorption are degradation products of bone collagen, but non collagenous proteins such as bone sialoprotein and tartrate-resistant acid phosphatase are also used as bone resorption marker.

Rai et al.; JPRI, 33(41A): 26-40, 2021; Article no.JPRI.72650

Hydroxyproline (Hyp) Constitutes 12-14% of the total amino acid content of mature collagens, but only 10% of Hyp released during bone resorption reaches the urine in free or peptide-bound forms. Urinary Hyp has long served as the only marker of bone resorption, despite the fact that significant amounts of urinary Hyp are derived from the degradation of newly synthesized collagens, from collagens of tissues other than bone, and from the diet. Today, Hyp is considered a nonspecific index of collagen turnover and has been largely replaced by more specific techniques. The hydroxylysineglycosides are integral parts of bone collagen and occur in two forms: glycosyl-galactosylhydroxylysine (Glc-Gal-Hyl) and galactosylhydroxylysine (Gal-Hyl). Both components are released into the circulation during collagen degradation and may be measured in urine by high-performance liquid chromatography (HPLC). The ratio of the two glycosides may allow for the recognition of tissue specificity. Although the hydroxylysine have potential as markers of bone resorption, their major disadvantage is presently the absence of a convenient immunoassay format. They are the first to the excreted in the urine during bone resorption [49].

The hydroxypyridinium crosslinks of collagen, pyridinoline (PYD) and deoxy pyridinoline (DPD), are formed during the extracellular maturation of fibrillar collagens and are released upon the degradation of mature collagens. The measurement of PYD and DPD is not influenced by the degradation of newly synthesized collagens and independent of dietary sources. In addition, the two components show a high specificity for skeletal tissues. While PYD is found in cartilage, bone, ligaments and vessels, DPD is found in bone and dentin only [50].

Both crosslink components may be measured by a reverse-phase ion-paired HPLC technique. In urine PYD and DPD are present both as free moieties (about 40%) and peptide-bound (about 60%). In addition, the free (non-peptide-bound) forms can be detected by direct immunoassays
(free DPD. Prilinks-D'). Sensitive (free DPD, 'Pyrilinks-D'). immunoassays are available for the measurement of type I collagen telopeptides in urine and serum. Currently, these include a serum radioimmunoassay (RIA) for the carboxyterminal type I collagen telo-peptide generated by matrix metalloproteases (CTX-MMP, also called 'ICTP') in serum, several immunoassays involving a syntheticoctapeptide from the C telopeptide of type I collagen containing the

crosslinking site (CTX-I, 'Cross laps') and an enzyme-linked immunosorbent assay (ELISA) for the cross-linked N-terminal telopeptide of type I collagen (NTX-I, 'Osteomark'). The pyridinium crosslinks and the collagen telopeptide involving the crosslinking site are currently considered the best indices for the assessment of bone resorption. Their urinary levels need to be corrected by creatinine excretion. However a study in Manipur claimed that DPD did not correlate well with DEXA and thus implying DPD as not a good indicator for osteoporosis [51].

Carboxy terminal crosslinked telopeptide (CTX- Beta Cross Lap): Cathepsin K degrades type I collagen of bone to form (CTX). They exist in two forms: α and β, in growing children the ratio is of the isoforms are altered. It is recommended as the marker for resorption by international osteoporosis foundation. They are altered in other metabolic disorders of bone [52]. However, there are studies that also claim CTX as not a good indicator for detecting osteoporosis.

Amino Terminal Crosslinked Telopeptides (Ntx): NTX like CTX are degraded from type 1 collagen by cleavage of N-terminal region by cathepsin K. This mechanism occurs during bone resorption. NTX is measured both in urine and serum using monoclonal antibody against specific N-terminal epitope.

Bone sialoprotein (BSP) accounts for 5-10% of the non-collagenous matrix of bone. The glycoprotein has been shown to be a major synthetic product of active osteoblasts and odontoblasts. BSP may play an important role in cell-matrix adhesion processes and in the supramolecular organization of the extracellular matrix of mineralized tissues. Immunoassays have been developed for the measurement of the immunoreactive form of BSP in serum. Based upon clinical data and the rapid reduction of
serum BSP levels following intravenous serum BSP levels following bisphosphonate treatment, it is assumed that serum BSP reflects processes mainly related to bone resorption, they are overexpressed in malignant tissues and its role in cancer management is well known but data are lacking to assess the clinical utility of this new marker in osteoporosis [53].

Tartrate-resistant acid phosphatase (TRACP) exists in two sub-isoforms named 5a and 5b, of which only TRACP-5b has been shown to be characteristic for osteoclasts. Recently, *Rai et al.; JPRI, 33(41A): 26-40, 2021; Article no.JPRI.72650*

immunoassays forTRACP-5b have been described and preliminary clinical results indicate that this marker may be useful to assess osteoclast activity [54].

9. RECENT MARKERS

Receptor activator nuclear factor kappa -B ligand: RANKL reflects the activity in bone osteoclasts in a microenvironment. The activity of RANKL and RANK is explained earlier and as they increase osteoclastic role and its mechanism as a resorption marker is still not well understood. These are currently under research setting to be proven as novel biomarkers of bone resorption in osteoporosis.

Osteoprotegerin: This marker also depicts bone cell mechanism. Unlike RANKL they are synthesized by osteoblasts and inhibit the binding of osteoclasts to RANK thus inhibiting bone resorption. This marker requires further analysis on its role in osteoporosis.

10. BIOLOGICAL VARIATIONS IN BONE TUMOR MARKERS

10.1 Uncontrollable Biological Variation

Renal Function: In the elderly gradual renal impairment may lead to an increase in osteocalcin and in other markers metabolized and/or excreted by the kidney (pyridinoline crosslinks and related peptides); care should be taken in the interpretation of these bone markers when the creatinine clearance decreases below 30 ml/min [55].

Pregnancy and lactation: Pregnancy and lactation place a considerable burden on the maternal skeleton to provide calcium for the growing fetus and infant. The greatest demand from the fetus for calcium comes in the third trimester, but an anticipatory mechanism results in a gradual increase in bone resorption from the sixteenth week of pregnancy onward, followed by an increase in bone formation [56]. Contrary to this overall pattern, serum levels of Osteocalcin appear to decrease and may even be undetectable during pregnancy. It has been suggested that this is due to placental clearance of Osteocalcin. However, it may also be due the type of assay used and to the increase in renal function during pregnancy [57]. At term, markers of bone resorption such as NTX, PINP and CTX increase and return to normal after delivery.

Rai et al.; JPRI, 33(41A): 26-40, 2021; Article no.JPRI.72650

Suggesting imprecision in BTM values during pregnancy.

Drugs: Corticosteroid treatment are misused thoroughly in India as they are freely available. They significantly reduce OC values [58]. Long term anticonvulsant therapy and GnRH agonist treatment both result in significant increases in markers of bone turnover. In contrast thiazide diuretics decrease bone turnover [59]. The effect of oral contraception (OCP) on bone turnover appears to be age-dependent. Intake of OCP in pre menopause has given a protective effect to bone thus reducing the bone turnover markers in post-menopausal women. Studies on the effect of oral contraception give inconsistent results [60].

Immobility: Bed rest results in a very rapid increase in markers of bone resorption. Urinary excretion of PYD and DPD are significantly increased after only 2 days and by 40% after a week. Markers of bone formation change little or remain unchanged during bed rest or immobility. In elderly, partially immobile subjects, the increase in urinary HYP is related to the degree of immobility. Once remobilization occurs resorption markers gradually return to initial levels, although paradoxically PICP may increase [61].

10.2 Controllable Sources of Biological Variability

Circadian Rhythm: Circadian variability has more impact on markers of bone turnover than most other sources of variability. Most markers of bone turnover are increased at night, reaching a peak between 0200 and 0800 hours, after which they decrease rapidly and reach a nadir between 1300 and 2300 hours. The amplitude of the rhythm is considerably greater for resorption markers than for formation markers. Serum OC are increased by 20% at night compared with their nadir in the early afternoon. CTX also observes circadian variation as they peak in the second half of night. 20% lower levels are observed post prandially. Hence fasting samples are preferred. Bone ALP has a somewhat different circadian rhythm with a peak between 1100 and 1400 hours and possibly another peak at 2330 hours. Calcium supplementation taken at night and bisphosphonate treatment can both suppress the circadian rhythm of markers of bone resorption [62].

Exercise: Exercise may affect the variability of markers of bone turnover in two ways: the effect of persistent exercise and the acute effect of a bout of exercise within a day of the sample collection. Sub-acute exercise results in an increase in bone formation markers and a decrease in bone resorption markers. In most studies the acute effect of exercise is to increase markers of collagen formation and degradation by 15-40%. These increases persist for 24 hr and possibly for as long as 72 hr. It is therefore important to enquire about regular exercise and ask the subject to refrain from exercise for at least 24 hr before samples are collected [63].

Diet: Serum and urinary levels of most markers of bone turnover are unaffected by diet, with the exception of HYP and CTX. Before samples are collected for HYP and CTX measurements, subjects must have an overnight fast. Specific dietary restrictions are therefore only applicable to HYP measurements.

Reference Ranges: Each laboratory should establish its own reference ranges. Age, gender, menopausal status and race all affect levels of markers of bone turnover. Therefore, separate reference ranges should be established for men, premenopausal women and postmenopausal women. Because markers are still elevated in the third decade the male and premenopausal reference ranges should only include subjects over the age of 30 years. Standardized time and conditions for sample collection must also be defined for each reference range.

11. ASSESSMENT OF BONE TURNOVER MARKERS IN OSTEOPOROSIS IN INDIA

Sachdeva.A et al observed an increased excretion of hydroxyproline in postmenopausal women as compared to the premenopausal women. This increase was statistically highly significant (p< 0.001) serum alkaline phosphatase and Calcium was used and was found to be significantly increased in postmenopausal women reflecting increased bone activity (osteoclastic and osteoblastic) as compared to premenopausal women. A study demonstrated significantly lower serum calcium level in the post-menopausal group, while the ALP level was significantly lower in premenopausal women suggesting increase in bone turnover with accelerated loss of bone mass in post-menopausal women. [64] Mean serum values of NTX increased significantly in post-menopausal women with increase in the duration of menopause and BMD values decreased and correlated well with NTX suggesting increased bone resorption in women with low BMD [65]. Bone specific ALP, deoxypyridinoline (DPD) and correlation of BMD was assessed in chronic kidney disease (CKD) patients in India. Osteoporotic subjects had higher BAP, and DPD, suggesting higher bone turnover in osteoporosis in CKD patients. 80% were having vitamin D deficiency and 13% had insufficiency [66]. A negative correlation of serum OC and BMD was seen in osteoporotic postmenopausal women, suggesting BTMs and BMD together are better predictors of osteoporotic fractures [67-70] However, Rai et al and gurban etal proposed low serum OC levels in osteoporotic post-menopausal women as compared to healthy post-menopausal woman, suggesting low bone formation is prone to fractures [71]. Serum BSP was significantly elevated in postmenopausal osteoporosis and positively correlated with NTX &other resorption markers as compared to healthy perimenopausal controls, suggesting elevated bone resorption markers in osteoporosis. Studies showed increased bone turnover markers in postmenopausal women as compared to premenopausal women, implying higher bone turnover immediately after menopause [72-74]. Similar results were seen with serum Osteoprotegerin in the previous age group.

A study in Sikkim proposed NTX and urinary HYP as markers in diagnosing osteoporosis in men whereas osteocalcin as an indicator in women. They concluded that bone turnover markers are able to predict low bone mass and will help to predict the risk of osteoporosis [75]. Abraham. S et al did a study on osteoporosis and osteopenia in India and found that osteopenia was more in the age group of 40 – 49 years and osteoporosis was more in the age group of 60 – 69 years [76].

12. CONCLUSION

Unfortunately, rural India still is lagging behind socio- economically and in health care, DEXA is not accessible to many living in India. Thus, biochemical assays were studied as they are affordable, non-invasive, not site specific, reflects the turnover of entire skeleton and can detect early changes in the bone. Thus, concluding that bone turnover markers act as important tools in the management of osteoporosis irrespective of the risk factors. Fracture risk can be predicted based on remodeling rates as epidemiological studies demonstrated them as independent

Rai et al.; JPRI, 33(41A): 26-40, 2021; Article no.JPRI.72650

contributors of fracture risk. Understanding the variations and availability of reliable, rapid and standardized turnover assays assists in better utilization of these markers in the management of osteoporosis.

CONSENT

Not applicable

ETHICAL APPROVAL

Not applicable

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Kanis JA, Cooper C, Rizzoli R, Reginster JY. Executive summary of European guidance for the diagnosis and management of osteoporosis in postmenopausal women. Aging clinical and experimental research. 2019;31(1):15- 7.
- 2. Riggs BL, Melton III LJ. The prevention and treatment of osteoporosis. New England Journal of Medicine. 1992;327(9): 620-7.
- 3. Rai M, Rai T, Rai S, D'Sa J, Parinita .K et al Serum osteocalcin: A potential biomarker for primary osteoporosis.
International Journal of Clinical International Journal of Clinical Biochemistry and Research. 2018;5(3): 392-396.
- 4. Khadilkar AV, Mandlik RM. Epidemiology and treatment of osteoporosis in women: an Indian perspective. International journal of women's health. 2015;7:841.
- 5. WHO Study Group. Assessment of fracture risk and its application to screening for postmenopausal osteoporosis. Report of a WHO Study Group. World Health Organ. Tech. Rep. Ser. 1994;843:1-29.
- 6. Khadilkar AV, Mandlik RM. Epidemiology and treatment of osteoporosis in women: an Indian perspective. International journal of women's health. 2015;7:841.
- 7. Kuo TR, Chen CH. Bone biomarker for the clinical assessment of osteoporosis: recent developments and future perspectives. Biomarker research. 2017;5(1):1-9.
- 8. Blumer MJ. Bone tissue and histological and molecular events during development

of the long bones. Annals of Anatomy-Anatomischer Anzeiger. 2021;16:151704.

- 9. Eswaran SK, Gupta A, Adams MF, Keaveny TM. Cortical and trabecular load sharing in the human vertebral body. Journal of Bone and Mineral Research. 2006;21(2):307-14.
- 10. Liu XS, Cohen A, Shane E, Stein E, Rogers H, Kokolus SL, Yin PT, McMahon DJ, Lappe JM, Recker RR, Guo XE. Individual trabeculae segmentation (ITS)– based morphological analysis of high-resolution peripheral quantitative computed tomography images detects abnormal trabecular plate and rod microarchitecture in premenopausal women with idiopathic osteoporosis. Journal of bone and mineral research. 2010;25(7):1496-505.
- 11. Demirtaş Ö, Demirtaş G, Hurşitoğlu BS, Terzi H, Şekerci Z, Ök N. Is grand multiparity a risk factor for osteoporosis in postmenopausal women of lower socioeconomic status?. European review for medical and pharmacological sciences. 2014;18(18):2709-14.
- 12. Grigoriadis AE, Heersche JN, Aubin JE. Differentiation of muscle, fat, cartilage, and bone from progenitor cells present in a bone-derived clonal cell population: effect of dexamethasone. The Journal of cell biology. 1988;106(6):2139-51.
- 13. Capulli M, Paone R, Rucci N. Osteoblast and osteocyte: games without frontiers. Archives of biochemistry and biophysics. 2014;561:3-12.
- 14. Fakhry M, Hamade E, Badran B, Buchet R, Magne D. Molecular mechanisms of mesenchymal stem cell differentiation towards osteoblasts. World journal of stem cells. 2013;5(4):136.
- 15. Jimi E, Hirata S, Osawa K, Terashita M, Kitamura C, Fukushima H. The current and future therapies of bone regeneration to repair bone defects. International journal of dentistry. 2012;2012.
- 16. Franz‐Odendaal TA, Hall BK, Witten PE. Buried alive: how osteoblasts become osteocytes. Developmental dynamics: an official publication of the American Association of Anatomists. 2006;235(1): 176-90.
- 17. Currey JD. The many adaptations of bone. Journal of biomechanics. 2003;36(10): 1487-95.
- 18. Schaffler MB, Cheung WY, Majeska R, Kennedy O. Osteocytes: master orchestr-

ators of bone. Calcified tissue international. 2014;94(1):5-24.

- 19. Boyce BF, Hughes DE, Wright KR, Xing L, Dai A. Recent advances in bone biology provide insight into the pathogenesis of bone diseases. Laboratory investigation; a journal of technical methods and pathology. 1999;79(2):83-94.
- 20. Sodek J, Mckee MD. Molecular and cellular biology of alveolar bone. Periodontology. 2000;24(1):99-126.
- 21. Miyamoto T. The dendritic cell-specific transmembrane protein DC-STAMP is essential for osteoclast fusion and osteoclast bone-resorbing activity. Modern rheumatology. 2006;16(6):341-2.
- 22. Pirro M, Leli C, Fabbriciani G, Manfredelli MR, Callarelli L, Bagaglia F, Scarponi AM, Mannarino E. Association between circulating osteoprogenitor cell numbers and bone mineral density in postmenopausal osteoporosis. Osteoporosis international. 2010;21(2): 297-306.
- 23. Miller SC, de Saint-Georges L, Bowman BM, Jee WS. Bone lining cells: structure and function. Scanning microscopy. 1989;3(3):27.
- 24. Mosley JR. Osteoporosis and bone functional adaptation: mechanobiological regulation of bone architecture in growing and adult bone, a review. Journal of rehabilitation research and development. 2000;37(2):189.
- 25. Aszódi A, Bateman JF, Gustafsson E, Boot-Handford R, Fässler R. Mammalian skeletogenesis and extracellular matrix: what can we learn from knockout mice?. Cell structure and function. 2000;25(2):73- 84.
- 26. Yagami K, Suh JY, Enomoto-Iwamoto M, Koyama E, Abrams WR, Shapiro IM, Pacifici M, Iwamoto M. Matrix GLA protein is a developmental regulator of chondrocyte mineralization and, when
constitutively expressed. blocks constitutively endochondral and intramembranous ossification in the limb. Journal of Cell Biology. 1999;147(5):1097-108.
- 27. Datta HK, Ng WF, Walker JA, Tuck SP, Varanasi SS. The cell biology of bone metabolism. Journal of clinical pathology. 2008;61(5):577-87.
- 28. Zimmerman D, Jin F, Leboy P, Hardy S, Damsky C. Impaired bone formation in transgenic mice resulting from altered integrin function in osteoblasts. Developmental biology. 2000;220(1):2-15.
- 29. Florencio-Silva R, Sasso GR, Sasso-Cerri E, Simões MJ, Cerri PS. Biology of bone tissue: structure, function, and factors that influence bone cells. BioMed research international; 2015.
- 30. Parker SE, Troisi R, Wise LA, Palmer JR, Titus-Ernstoff L, Strohsnitter WC, et al. Menarche, menopause, years of menstruation, and the incidence of osteoporosis: the influence of prenatal exposure to diethylstilbestrol. The Journal of Clinical Endocrinology & Metabolism. 2014;99(2):594-601.
- 31. Pothiwala P, Evans EM, Chapman-Novakofski KM. Ethnic variation in risk for osteoporosis among women: a review of biological and behavioral factors. Journal of women's health. 2006;15(6):709-19.
- 32. Mitra S, Desai M, Khatkhatay MI. Vitamin D receptor gene polymorphisms and bone mineral density in postmenopausal Indian women. Maturitas. 2006;55(1):27-35.
- 33. Rai T, Rai M, Dsa J, Rai S, Sushith P, Bhandary R. Impact of Seasonal Variation in Association with Other Factors on Vitamin D Status among Mangalorean Population. Journal of Evolution of Medical and Dental Sciences. 2021;10(9):589-95.
- 34. Maïmoun L, Sultan C. Effects of physical activity on bone remodeling. Metabolism. 2011;60(3):373-88.
- 35. Ward KD, Klesges RC. A meta-analysis of the effects of cigarette smoking on bone mineral density. Calcified tissue international. 2001;68(5):259-70.
- 36. Buckley L, Humphrey MB. Glucocorticoidinduced osteoporosis. New England Journal of Medicine. 2018;379(26):2547- 56.
- 37. Gurban CV, Balas MO, Vlad MM, Caraba AE, Jianu AM, Bernad ES, et al. Bone turnover markers in postmenopausal osteoporosis and their correlation with bone mineral density and menopause duration. Romanian J Morphol Embryol. 2019;60(4):1127-35.
- 38. Marwaha RK, Tandon N, Gupta Y, Bhadra K, Narang A, Mani K, et al. The prevalence of and risk factors for radiographic vertebral fractures in older Indian women and men: Delhi Vertebral Osteoporosis Study (DeVOS). Archives of osteoporosis. 2012;7(1):201-7.
- 39. Rath S, Yadav L, Tewari A, Chantler T, Woodward M, Kotwal P, et al. Management of older adults with hip fractures in India: a mixed methods study

of current practice, barriers and facilitators, with recommendations to improve care pathways. Archives of osteoporosis. 2017;12(1):1-3.

- 40. Caracchini G, Cavalli L. Severe
osteoporosis: Diagnosis of femoral osteoporosis: fractures. Clinical cases in mineral and bone metabolism. 2010;7(2):97.
- 41. Bhandari M, Schemitsch EH. Fragility fractures: A global problem?.- radius fracture
- 42. Ozaki A, Tanimoto T, Yamagishi E, Sato S, Tsukada M, Sawano T, et al. Finger fractures as an early manifestation of primary hyperparathyroidism among young patients: a case report of a 30-year-old male with recurrent osteoporotic fractures. Medicine. 2016;95(20).
- 43. Mithal A, Bansal B, Kyer CS, Ebeling P. The Asia-pacific regional auditepidemiology, costs, and burden of osteoporosis in India 2013: a report of international osteoporosis foundation. Indian journal of endocrinology and metabolism. 2014;18(4):449.
- 44. Qu XL, Zheng B, Chen TY, Cao ZR, Qu B, Jiang T. Bone Turnover Markers and Bone Mineral Density to Predict Osteoporotic Fractures in Older Women: A Retrospective Comparative Study. Orthopaedic surgery. 2020;12(1):116-23.
- 45. Silambanan S, Menon PG, Arunalatha P. Comparison of bone mineral density with biochemical parameters and prevalence of osteopenia and osteoporosis in South
Indian population. Biomedical and Indian population. Biomedical and Pharmacology Journal. 2018;11(4):2209- 14.
- 46. Zanatta LC, Boguszewski CL, Borba VZ, Moreira CA. Association between undercarboxylated osteocalcin, bone mineral density, and metabolic parameters in postmenopausal women. Archives of endocrinology and metabolism. 2018;62: 446-51.
- 47. Hatkar SS, Kadam SS, Khatkhatay MI, Desai MP. Screening and Assessment of Bone Health in Indian Women Using an Indigenous ELISA of Human Osteocalcin a Bone Turnover Marker. Indian Journal of Clinical Biochemistry. 2020;35(4):436-41.
- 48. Pal R, Aggarwal A, Sachdeva N, Ram S, Garg A, Bhansali A, et al. Age-and sexspecific concentrations of bone remodeling markers in healthy Indian adults with and without vitamin D deficiency. Archives of Osteoporosis. 2021;16(1):1-1.

Rai et al.; JPRI, 33(41A): 26-40, 2021; Article no.JPRI.72650

- 49. Sachdeva A, Seth S, Khosla AH, Sachdeva S. Study of some common biochemical bone turnover markers in postmenopausal women. Indian journal of clinical biochemistry. 2005;20(1):131-4.
- 50. Robins SP, Woitge H, Hesley R, Ju J, Seyedin S, Seibel MJ. Direct, enzyme‐linked immunoassay for urinary deoxypyridinoline as a specific marker for measuring bone resorption. Journal of Bone and Mineral Research. 1994;9(10): 1643-9.
- 51. Rajkumari K, Singh AJ, Khanna T, Singh AB. Normative data of the urinary bone resorption biomarkers for osteoporosis DPD, CTx and NTx among females in Manipur and its correlation with bone mineral density measured by DEXA scan.
- 52. Garnero P, Fledelius C, Gineyts E, Serre CM, Vignot E, Delmas PD. Decreased β‐Isomerization of the C‐Terminal Telopeptide of Type I Collagen α1 Chain in Paget's Disease of Bone. Journal of bone and mineral research. 1997;12(9):1407-15.
- 53. Karadag A, Ogbureke KU, Fedarko NS, Fisher LW. Bone sialoprotein, matrix metalloproteinase 2, and αvβ3 integrin in osteotropic cancer cell invasion. Journal of the National Cancer Institute. 2004;96(12): 956-65.
- 54. Halleen JM, Ylipahkala H, Alatalo SL, Janckila AJ, Heikkinen JE, Suominen H, et al. Serum tartrate-resistant acid phosphatase 5b, but not 5a, correlates with other markers of bone turnover and bone mineral density. Calcified tissue international. 2002;71(1).
- 55. Nishizawa Y, Miura M, Ichimura S, Inaba M, Imanishi Y, Shiraki M, et al. Executive summary of the Japan osteoporosis society guide for the use of bone turnover markers in the diagnosis and treatment of osteoporosis (2018 Edition). Clinica Chimica Acta. 2019;498:101-7.
- 56. Essley B, McNanley T, Cooper B, McIntyre A, Witter F, Harris Z, et al. Osteoprotegerin in pregnant adolescents differs by race and is related to infant birth weight z-score. Journal of developmental origins of health and disease. 2011;2(5):272-9.
- 57. Cole DE, Gundberg CM, Stirk LJ, Atkinson SA, Hanley DA, Ayer LM, et al. Changing
osteocalcin concentrations during osteocalcin concentrations pregnancy and lactation: implications for maternal mineral metabolism. The Journal of Clinical Endocrinology & Metabolism. 1987;65(2):290-4.
- 58. Dua A, Das P, Ravindran V. Glucocorticoids: A review of its adverse effects including bone loss. Indian Journal of Rheumatology. 2019;14(5):90.
- 59. Feldkamp J, Becker A, Witte OW, Scharff
D. Scherbaum WA. Long-term Long-term anticonvulsant therapy leads to low bone mineral density—evidence for direct drug effects of phenytoin and carbamazepine on human osteoblast-like cells. Experimental and clinical endocrinology & diabetes. 2000;108(01):37-43.
- 60. Corson SL. Oral contraceptives for the prevention of osteoporosis. The Journal of reproductive medicine. 1993;38(12 Suppl): 1015-20.
- 61. Chen JS, Cameron ID, Cumming RG, Lord SR, March LM, Sambrook PN, et al. Effect of age‐related chronic immobility on markers of bone turnover. Journal of Bone and Mineral Research. 2006;21(2):324-31.
- 62. Shetty S, Kapoor N, Bondu JD, Thomas N, Paul TV. Bone turnover markers: Emerging tool in the management of osteoporosis. Indian journal of endocrinology and metabolism. 2016;20(6):846.
- 63. Smith C, Tacey A, Mesinovic J, Scott D, Lin X, Brennan-Speranza TC, et al. The effects of acute exercise on bone turnover markers in middle-aged and older adults: A systematic review. Bone. 2020:115766.
- 64. Bhattarai T, Bhattacharya K, Chaudhuri P, Sengupta P. Correlation of common biochemical markers for bone turnover, serum calcium, and alkaline phosphatase
in post-menopausal women. The in post-menopausal women. The Malaysian journal of medical sciences: MJMS. 2014;21(1):58.
- 65. Kumar A, Devi SG, Mittal S, Shukla DK, Sharma S. A hospital based study of biochemical markers of bone turnovers & bone mineral density in north Indian women. The Indian journal of medical research. 2013;137(1):48.
- 66. Jabbar Z, Aggarwal PK, Chandel N, Khandelwal N, Kohli HS, Sakhuja V, et al. Noninvasive assessment of bone health in Indian patients with chronic kidney disease. Indian journal of nephrology. 2013;23(3):161.
- 67. Kalaiselvi VS, Prabhu K, Mani Ramesh VV. The association of serum osteocalcin with the bone mineral density in postmenopausal women. Journal of clinical and diagnostic research: JCDR. 2013;7(5):814.
- 68. Fernandez C, Jayasingh P, Pandi S, Solaiyappan N, Murugesan A. Association

Rai et al.; JPRI, 33(41A): 26-40, 2021; Article no.JPRI.72650

of bone mineral density (BMD), body mass index (BMI) and serum osteocalcin in South Indian post-menopausal women. Indian Journal of Medical Specialities. 2018 ;9(2):52-5.

- 69. Singh S, Kumar D, Lal AK. Serum osteocalcin as a diagnostic biomarker for primary osteoporosis in women. Journal of clinical and diagnostic research: JCDR. 2015;9(8):RC04.
- 70. Jagtap VR, Ganu JV, Nagane NS. BMD and serum intact osteocalcin in postmenopausal osteoporosis women. Indian Journal of Clinical Biochemistry. 2011;26(1):70-3.
- 71. Rai M, Rai T, DSA J, Rai S. Bone Turnover Markers; an Emerging Tool to Detect Primary Osteoporosis. Journal of Clinical & Diagnostic Research. 2018;12(12).
- 72. Rai T, Rai M, Jeppu A K, Shetty M, Rai S et al. Effect of menopause on bone turnover. World Journal of Pharmaceutical Sciences. 2(8):775-779.
- 73. Indumati V, Patil VS, Jailkhani R. Hospital based preliminary study on osteoporosis in postmenopausal women. Indian Journal of clinical biochemistry. 2007;22(2) :96.
- 74. Jharna S, Nupur S, Purnima DS, Angoorbala B. Assessment of bone loss in postmenopausal women by evaluation of urinary hydroxyproline and serum status of osteocalcin. Int. Res. J. Biol. Sci. 2013;2: 11-4.
- 75. Soibam D, Singh T P, Nandy P, Baruah A. Role of biochemical markers in the prediction of osteopenia and osteoporosis in men and women of Sikkim, India. International Journal of Research in Medical Science. 2018;6(12):4077.
- 76. Babu AS, Ikbal FM, Noone MS, Joseph AN, Samuel P. Osteoporosis and osteopenia in India: A few more observations. Indian Journal of Medical Sciences. 2009;63(2):76-77.

© 2021 Rai et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

> *Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle4.com/review-history/72650*