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Influence of Silver Nanoparticles on Onion (*Allium cepa* L.) Seed Germination and Seedling Vigor

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

During this study on the influence of silver nanoparticles (AgNPs) on growth of L-883 onion varieties $Allium\ cepa\ L$. AgNPs 100 nm size were used in four different concentrations (25, 50, 75, and 100 ppm) and T_0 was kept as control. The seeds of onion were brought from NHRDF Indore. The findings revealed that 25 ppm of silver nanoparticle led to the highest seed germination percentage as compared to other concentrations. On the other hand, all the other concentrations were found to have a harmful effect on both seed germination and seedling growth as the level of toxicity are strongly associated with concentration of the suspensions. AgNPs of 100ppm concentration was

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found to have lowest rate of seed germination, however non-significant difference was found in 75ppm and 100ppm concentrations. Therefore, a concentration of 25ppm of AgNPs proved to be the most effective treatment for promoting optimal seed germination and seedling growth in onions.

Keywords: Onion; nanoparticles; seed germination; seedling growth.

1. INTRODUCTION

Onion (Allium cepa) is an important bulb crop and a key vegetable in India. It belongs to Alliaceae family which includes several more economically substantial crops. onions are the second most extensily consumed vegetable in the world and they are the oldest to be produced [1].

Onion has anti-bacterial, anti-fungal properties and many medicinal properties such as anti-cancer properties and work as an anti-cholesterol agent. Onion also has an antioxidant component quercetin in it [2] In add up to trade of vegetables in India, onions have the biggest section of Ledger account of 76.2 per cent yet its efficiency is lower than numerous of the nation. Insufficiency of high-quality onion's germplasm is one of the main reasons for less yield [3-5]. Seed quality parameters such as seed size and seed weight also influence the ultimate reduction in the yield of onion [6].

In expansion, it has been detailed to be wealthy in phytochemicals particularly flavanols which has therapeutic esteem [7]. Within the World, India ranks second after china in terms of range and production [8,9].

During storage, onion seed rapidly misfortune its reasonability because they generate free radicals by lipid peroxidation which result in low germination rate. As Current technologies are insufficiently effective in tackling these issues on a large scale. nevertheless, nanotechnologies hold promise for finding a solution. Nanoparticles appear diverse impacts on seed germination of distinctive plants [10-13].

According to Noshad et al. [14], AgNPs dramatically boosted *Solanum lycopersicum* seed germination and seedling growth. In tomato $AgNP_s$ increased the length and quantity of roots in seedlings, as well as the amount of nitrogen and phosphorus in the leaves [15].

2. MATERIALS AND METHODS

AgNPs with different concentration was used in this experiment. The seeds of onion (L-883) was brought from National Horticultural Research and

Development Foundation (NHRDF) Madhya Pradesh. AgNPs of strengths of 25 ppm (T_1) , 50 ppm (T_2) , 75 ppm (T_3) , and 100 ppm (T_4) were prepared in distilled water using a water bath as treatments.

2.1 Procedure

The experiment was conducted in completely randomized design (CRD) [16] with three replicates. For each treatment, 50 germplasm were placed in each replicate. To ensure surface sterility, seeds were soaked in 5% sodium hypochlorite solution for 20 minutes [17] and subsequently washed many times with distilled water. 150 seeds were submerged in the solution containing AgNPs for around 3 hours. As supplementary control, seeds were kept in distilled water for the same period. The vessel was frequently shaken for uniform adsorption of AgNPs. Whatman no. 1 filter paper was placed on the Petri plate with a diameter of 100 x 15 mm and 2.5 ml of solution was poured onto it, and in control 2.5 ml of double-distilled water was poured. The Petri dishes were then placed in a germination chamber at 26 ± 5 °C and 96 ± 2% relative humidity. Germination were recorded daily and further observations were taken on 6th days, and the last observation was recorded on the 14th day.

Parameters measured in the study:

Germination Percentage = $(Gsn^{-1}) \times 100$

where;

Gs = number of germinated germplasm n = Total number of seed used in the test.

By counting the cumulative germination against each day Cumulative germination percentage is calculated.

2.2 Germination rate index (GSI)

The germination index was calculated as the total number of germplasm germinated per day divided by the number of days from sowing to germination, according to the Maguire equation [18].

2.3 Mean Germination Time (MGT)

"Mean germination time was calculated by the formula given by [19]

 $MGT = n1 \times d1 + n2 \times d2 + n3 \times d3 + -----/$ Total number of days"

"where:

n= Number of seed which have germinated d = Number of days

2.4 Mean Daily Germination (MDG)

Mean daily germination was calculated by the formula given by [18]

MDG = Number of germinated seeds / Total number of days

2.5 Vigour Index

The vigour index was calculated using the approach recommended by Anderson [20].

Vigour index = Germination percentage × Seedling length.

Tolerance Index (TI) = Average root length in AgNPs solution divided by longest root length in solution containing double distilled water x 100. [21]"

2.6 Root and Shoot Length

"The measurement of root length from the base of the collar to the tip of a root was done for all

normal seedlings, and the average value was in centimetres. The average value for the shoot length of seedlings from the germination test was measured on the 6th and 14th days, respectively, from around the collar area to the tip of the shot. A similar approach was taken to determine root-to-shoot ratios. Fresh mass was weighed with an electronic balance and was expressed in mg. seedlings were placed in an oven at a temperature of 65°C until the weight became constant and the weight was expressed in mg."

3. RESULTS AND DISCUSSION

"In the study, there was a significant increase in germination percentage (GP) in T1 (25 ppm AgNPs) compared to other treatments (Table 1). lowest germination % was observed in 100ppm of AgNPs in both the observations as high concentration of silver nanoparticle effects germination %. Germination speed index was observed maximum in 25ppm whereas T3 and T4 was at par but significantly lesser than control (Table 1). A cumulative germination percentage (CGP) means that germination has started on 3th day for all treatments except in T4 (100 ppm) where, the first germination was observed on 4th day. From the cumulative germination rates, it can be concluded that T2, T3, and T4 were partially toxic to the seedlings. It may also notice that seed of T₂ could manage themselves to pick the germination after 7 days. whereas, mean germination time was non-significant among all the treatment."

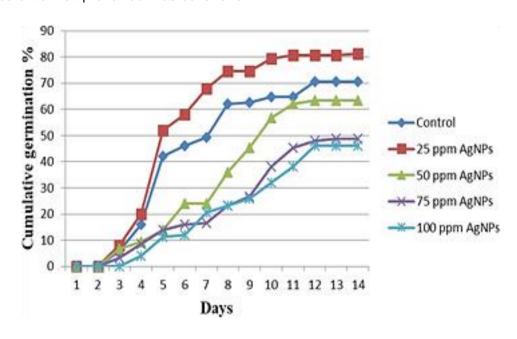


Fig. 1. Cumulative germination percentage of germplasm treated with AgNPs

Table 1. Germination and vigour parameters of onion

| Treatment | atment Germination % | | Germination speed Index | Mean germination time | Mean daily germination | | Vigor index | |
|------------|----------------------|----------------------|-------------------------|-----------------------|------------------------|----------------------|---------------------|----------------------|
| | 6 th day | 14 th day | - | | 6 th day | 14 th day | 6 th day | 14 th day |
| Control | 46.00 ^b | 70.66 ^b | 6.60 ^b | 2.23 | 3.83 ^b | 2.52 ^b | 155.34b | 641.35 ^b |
| T1 | 58.00a | 81.33a | 7.98 ^a | 2.36 | 4.83 ^a | 2.90a | 235.86a | 871.16a |
| T2 | 24.00 ^c | 66.00 ^b | 4.87 ^c | 2.46 | 2.00 ^c | 2.26 ^c | 62.48 ^c | 500.78c |
| T3 | 16.00 ^d | 50.00° | 3.6c ^d | 2.03 | 1.33 ^d | 1.73 ^d | 17.07 ^d | 250.42 ^d |
| T4 | 12.00 ^d | 46.00° | 3.12 ^d | 1.99 | 1.00 ^d | 1.64 ^d | 7.25 ^d | 216.42e |
| CD at 0.05 | 6.28 | 5.04 | 1.82 | NS | 0.53 | 0.20 | 19.97 | 28.75 |
| SEm | 2.03 | 1.62 | 0.27 | NS | 0.15 | 0.05 | 6.34 | 9.10 |

Table 2. Performance of onion seed for tolerance index, shoot and root ratio, seedling fresh weight and seedling dry weight

| Treatment | Tolerance index | | Shoot and root ratio (cm) | | Seedling fresh weight (mg) | Seedling dry weight(mg) | |
|-----------|---------------------|----------------------|---------------------------|----------------------|----------------------------|-------------------------|--|
| | 6 th day | 14 th day | 6 th day | 14 th day | _ | | |
| Control | 100.00b | 100.00 | 2.46a | 3.72 ^b | 20.27 ^b | 1.50 | |
| T1 | 140.61a | 120.57 | 1.97 ^b | 3.64 ^b | 25.89 ^a | 1.71 ^a | |
| T2 | 105.85 ^b | 94.47 | 1.50 ^c | 3.51 ^b | 21.40 ^b | 1.09 ^d | |
| T3 | 76.39 ^c | 51.90 | 0.45 ^d | 5.04 ^a | 16.47 ^c | 1.52 ^b | |
| T4 | 58.13 ^d | 52.37 | 0.27^{d} | 4.67 ^a | 17.40° | 1.30° | |
| CD (0.05) | 11.68 | 10.30 | 0.26 | 0.61c | 1.28 | 0.16 | |
| SEm | 3.26 | 3.22 | 0.3 | 0.06 | 0.5 | 0.04 | |

Table 3. Mean performance of onion seed for shoot length and root length.

| Treatment | Shoo | t length (mm) | Root length (mm) | | |
|-----------|---------------------|----------------------|---------------------|----------------------|--|
| | 6 th day | 14 th day | 6 th day | 14 th day | |
| Control | 24.00 b | 71.54 b | 9.77b | 19.213 ^b | |
| T1 | 27.03 a | 83.91a | 13.72a | 23.187a | |
| T2 | 15.59 ^b | 63.52c | 10.33 ^b | 18.133 ^b | |
| T3 | 3.37 ^c | 49.73d | 7.43 ^c | 9.947⁰ | |
| T4 | 1.48 ^d | 46.84 ^e | 5.70 ^d | 10.06 ^c | |
| CD (0.05) | 2.08 | 2.59 | 1.19 | 2.12 | |
| SEm | 0.68 | 0.84 | 0.37 | 0.69 | |

"The average daily germination was higher on days 6 and 14, but the average daily germination on day 6 was higher (4.83 %) than on day 14 (2.90). This means that 25 ppm AqNPs stimulate early germination and data represents that 50 ppm, 75 ppm, and 100 ppm AgNPs of populate early germination (Table 1). Treatment with 25 ppm AgNPs showed the maximum vitality index (VI) on days 6 and 14, which was significantly higher as compared to control. It was that vigour index gradually decreases as concentration increases as 50, 75 and 100ppm has less vigour index. Lowes vigour index was observed in T4 (Table 1). AgNPs incorporate with seed coats and helps in water uptake inside seeds which promotes starch metabolism and germination. It is also applied as a Nano-priming agent to enhance rice-aged seeds and starch metabolism [22]"

It was also noticed that treatment T2, T3 and T4 had very little VI compare to T₁ or control on 6th day, which indicates that in initial days there was minimum growth. Maximum tolerance index (TI) was observed in T₁ and minimum was observed in T₄ (Table 2). The decline in TI and VI was brought on by a higher proportion of seed germination inhibition, which was followed by a decrease in shoot height and root length as a result of the combined negative impacts of metal poisoning from AgNPs via oxidative stress. The treatment T1 had the highest seedling fresh weight (25.89 mg) compared to all other treatments, while T3 the lowest had seedling fresh weight (however it was comparable to T4).

Similarly, highest dry weight was recorded in T_1 and lowest dry weight was recorded in T_2 . It was notice that shoot length was less in T_4 and T_3 compare to other treatment (Table 3). maximum shoot length was observed in T_1 . Maximum and minimum root length was observed in T_1 and T_3 respectively (Table 3.) Pandey et al. [23] reported that root and shoot growths are more

affected by the Ag treatment than seed germination. The roots length was retarded to 1% and 0.5% in Oryza sativa and Brassica campestris; Vigna radiata, respectively when AG nanoparticles were used at higher concentration. Mazumdar et al. [24]

4. CONCLUSION

All parameters indicate that AgNPs at lower concentrations (25 ppm) will promote germination and have a positive effect on physiological activities, but higher concentrations of AgNPs are toxic for seedlings. Among all treatments, T₄ was most toxic to seedlings. 100 ppm AgNPs significantly reduced all parameters. The toxic effect of AgNPs is may be due to increased production of ROS during interaction between nanoparticle and plant [25]. Less concentration induced plantlets while higher concentration caused inhibition [26] AgNPs at 0-, 10-, 20-, and 30-mM increased plant growth and maintains the ionic balance of cells (Na+, K+, and Na+/K+ ratio) by decreasing the uptake of Na and CI and oxidative stress by salt-stressed plants [22].

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative Al technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

 Benitez V, Molla E, Martin-Cabrejas MA, Aguilera Y, Lopez-Andreu FJ, Esteban RM. Effect of sterilization on dietary fiber

- and physicochemical properties of onion by-products. Food Chem. 2011;127(2): 501–507.
- DOI: 10.1016/j.foodchem.2011.01.031
- Baghizadeh A, Baniasadi F, Bonjar GHS, Sirchi GRS, Massumi H, Jorjandi M Biocontrol of Botrytis allii Munn the causal agent of neck rot, the post-harvest disease in onion, by use of a new Iranian isolate of Streptomyces. American Journal of Agriculture and Biological Sciences. 2009; 4:72-78.
- Lee WM, Kwak JI, An YJ. Effect of silver nanoparticles in crop plants Phaseolus radiatus and Sorghum bicolor: media effect on phytotoxicity. Chemosphere. 2012;86: 491
- 4. Maguire JD. Speed of germination-aid in selection and evaluation for seedling emergence and vigour, Crop Sci. 1962;2: 176-177
- 5. Monica BC, Cremonini R. Nanoparticles and higher plants. Caryologia. 2009;62: 161-165
- Gamiely S, Randle, William, Mills, Harr, Smittle DA et al. Onion Plant Growth, Bulb Quality, and Water Uptake following Ammonium and Nitrate Nutrition. Hort Science: a publication of the American Society for Horticultural Science. 1991;26.
- Javadzadeh A, Ghorbanihaghjo A, Bonyadi S, Rashidi MR, Mesgari M, Rashtchizadeh Preventive effect of onion juice on seleniteinduced experimental cataract. Indian Journal of Ophthalmology. 2009;57:185-189.
- 8. Yin L, Cheng Y, Espinasse B, Colman PB, Auffan M, Wiesner M The effects of silver nanoparticles on *Lolium multiflorum*. Environ. Sci. Technol. 2011; 45:2360.
- Das R. Effect of silver Nano-particle on seed germination and seedling vigour of onion (Allium cepa L.). IJCS. 2018;6(3): 427-30.
- 10. Rico CM, Majumdar S, Gardea MD, Peralta Videa JR, Jorge L, Torresdey GL. Interaction of nanoparticles with edible plants and their possible implications in the food chain. J Agric. Food Chem. 2011; 59(8):3485-98.
- 11. Asharani PV, Wu YL, Gong Z, Valiyaveettil S. Toxicity of silver nanoparticles in zebrafish models. Nanotechnology. 2008; 19(25):255102.
- 12. ISTA. International Rules of Seed Testing. Seed Sci. & Technol. 2005;27:27-32

- Kumari M, Mukherjee A, Chandrasekaran N. Genotoxicity of silver nanoparticles in Allium cepa. Sci Total Environ. 2009;407: 5243-5246
- 14. Noshad A, Hetherington, Iqbal M. Impact of AgNPs on seed germination and seedling growth: a focus study on its antibacterial potential against *Clavibacter michiganensis* subsp. michiganensis infection in Solanum lycopersicum; 2019.
- 15. Guzman-Baez GA, Trejo-Tellez LI, Ramírez-Olvera SM, Salinas Ruíz J, BelloBello JJ, Alcantar-Gonzalez G, Hidalgo-Contreras JV, Gomez-Merino ´ FC. Silver nanoparticles increase nitrogen, phosphorus, and potassium; 2021.
- 16. Maxwell SC. Completely randomized design. Encyclopedia of Statistics in Behavioral Science; 2005.
- USEPA. Ecological effects test guidelines: Seed germination/root elongation toxicity test, OPPTS 850, 4200, EPA 712-C-96-154, Washington DC, 1996.
- Czabator FJ. Germination value: An index combining speed and completeness of pine seed germination. Forest Science. 1962;8:386-395.
- Ellis RH, Roberts EH. The quantification of ageing and survival in orthodox germplasm. Seed Sci. Tech. 1981;9:373-409
- Abdul-Baki AA, Anderson JD. Vigour deterioration of soybean germplasm by multiple criteria. Crop Sci. 1973;13:630-633.
- Saeed KM, Reza H, Fatemeh R, Solmaz N. Effect of Chemical Synthesis Silver Nanoparticles on Germination Indices and Seedlings Growth in Seven Varieties of Lycopersicon esculentum Mill (tomato). Plants. J Clust Sci. 2016;27:327-340.
- 22. Khan S, Zahoor M, Khan R, Ikram M Islam. The impact of silver nanoparticles on the growth of plants: The agriculture applications; 2023
- 23. Pandey C, Khan E, Mishra A, Sardar M, Gupta M. Silver nanoparticles and its effect on seed germination and physiology in *Brassica juncea* L. (Indian mustard) plant. Adv. Sci. Lett. 2014;20:1673–1676.
- Mazumdar H, Ahmed GU. Synthesis of silver nanoparticles and its adverse effect on seed germinations in oryza sativa, vigna radiata and Brassica campestris. Int. J. Adv. Biotechnol. Res. 2011;2:404–413.
- 25. Marslin G, Sheeba CJ, Franklin G. Nanoparticles alter secondary metabolism

in plants via ros burst. Front Plant Sci. 2017:8:832.

26. Mandal D, Bolander ME, Mukhopadhyay D, Sarkar G., Mukherjee P. The use of

microorganisms for the formation of metal nanoparticles and teir application. Appl. Microbiol. Biotechnol. 2006;69:485–492. DOI: 10.1007/s00253-005-0179-3.

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