



# Sequential Administration of Febuxostat, Amlodipine and Vitamin E Attenuate Oxidative Stress and Improve Spermatogenesis in Testicular Ischemia Reperfusion Injury in Wistar Rats

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## Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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## ABSTRACT

This study investigated the effects of sequential administration of febuxostat, amlodipine, and vitamin E on oxidative stress and spermatogenesis following testicular ischemia-reperfusion injury (TIRI) in Wistar rats. Ninety male rats (120-150 g) were divided into 9 groups (n=10 each): sham operation (SO), torsion-detorsion (TD), and seven treatment groups receiving different combinations of the drugs. The treatment groups included torsion + febuxostat + detorsion (TFD), torsion + detorsion + amlodipine (TDA), torsion + detorsion + vitamin E (TDV), and combinations thereof (TFDA, TFDV, TDAV, TFDVAV). TIRI was induced by 720° clockwise testicular torsion for 1 hour followed by detorsion. Febuxostat (5 mg/kg) was administered 30 minutes after torsion, amlodipine (2.5 mg/kg) immediately upon detorsion, and vitamin E (10 mg/kg) 30 minutes after detorsion.

Rats were sacrificed 56 days post-reperfusion. Testicular tissue was analyzed for antioxidant enzymes (superoxide dismutase, catalase), lipid peroxidation (malondialdehyde), total protein, inflammatory markers (serum nitrite, IL-1 $\beta$ ), and spermatogenesis indices (testicular biopsy score, Leydig cell count).

Results showed that TIRI significantly decreased antioxidant enzymes, significantly increased lipid peroxidation and inflammatory markers, and impaired spermatogenesis. In the TD group, superoxide dismutase (SOD) and catalase (CAT) activities were significantly decreased, while malondialdehyde (MDA) increased significantly compared to the SO group ( $p < 0.01$ ). Serum nitrite and IL-1 $\beta$  levels in the TD group were also increased ( $p < 0.001$ ). Furthermore, the testicular biopsy score and Leydig cell count in the TD group were significantly decreased in this study ( $p < 0.01$ ).

Sequential administration of febuxostat, amlodipine and vitamin E, particularly when all three were used (TFDAV group), significantly attenuated these changes. In the TFDAV group, SOD and CAT activities were improved, while MDA decreased compared to the TD group ( $p < 0.05$ ). Serum nitrite and IL-1 $\beta$  levels in the TFDAV group were also decreased ( $p < 0.001$ ). In addition, the testicular biopsy score and Leydig cell count in the TFDAV group increased in comparison with the TD group ( $p < 0.001$ ).

The study concludes that this sequential multi-drug approach shows promise in mitigating the long-term detrimental effects of TIRI on testicular function and fertility. The combination of febuxostat (a xanthine oxidase inhibitor), amlodipine (a calcium channel blocker), and vitamin E (an antioxidant) appears to provide protection against oxidative stress and inflammation induced by TIRI, thereby preserving spermatogenesis.

*Keywords: Oxidative stress; antioxidant enzymes; spermatogenesis; inflammation; lipid peroxidation.*

## 1. INTRODUCTION

Testicular ischemia-reperfusion injury (TIRI) is the primary pathophysiological event in testicular torsion repair [1]. It is caused by reoxygenation of the testis after episode of ischemia [2]. It is a common urological emergency in males of all ages [3]. TIRI has been reported to cause male infertility via activation of oxidative stress [4] which occur as a result of imbalance between pro-oxidants and antioxidants thereby resulting in cell or tissue injury [5]. Oxidative stress has been documented to disrupt the capacity of the germinal epithelium to differentiate into normal sperm cells [6].

Based on previous studies, TIRI produced copious amount of reactive oxygen species (ROS) which induced testicular damage via two

mechanisms; direct action on testicular components such as DNA, protein, lipids and carbohydrate [6] and indirect action via non-radical oxidants such as hydrogen peroxide, modulator via molecular bond, oxidative or nitrosative principle regulatory proteins. In addition to this, TIRI-induced oxidative stress has been reported to increase lipid peroxidation process, deplete antioxidant enzymes, activate oxido-inflammatory response and increase the rate of mitochondria mediated apoptosis in germ cells [7].

During TIRI, reactive oxygen species (ROS) is generated during the ischemic phase, early phase of reperfusion and minutes after reperfusion [8]. In the ischemic phase, ROS is generated via xanthine oxidase (XO) build-up, in the early phase of reperfusion via calcium-mediated ROS-production while minutes after

reperfusion ROS burst is generated by leukocyte recruitment to the site of injury [9]. This generated ROS is capable of forming peroxy radicals which can interfere with cellular structures such as proteins, lipids and DNA to cause severe oxidative damage to the testes [10,6,11]. The ROS generated minutes after reperfusion is capable of activating the release of inflammatory marker like interleukin-1beta from the resident macrophages and dendritic cells into the blood [12,13] which will accelerate the recruitment of neutrophils from the blood to the site of injury (testes) to trigger germ cell apoptosis thereby resulting in late organ damage in the long-run [8].

Based on the multifactorial nature of TIRI pathway before and after repair of torsion of the testes [6], it is necessary to prevent TIRI-induced oxidative stress damage to the testes urgently by blocking sources of ROS in the ischemic phase, early phase of reperfusion and minutes after reperfusion.

In this study, three major points are regarded as sources of testicular damage after testicular torsion onset and repair. Firstly, there is build-up of XO enzymes arising from adenosine triphosphate (ATP) depletion in the ischemic phase of testicular torsion which generate ROS that trigger oxidative stress [14,15]. This pathway was blocked by febuxostat to reduce oxidative stress induced testicular damage [16] in the ischemic phase. Secondly, there is activation of calcium-mediated ROS-induced apoptotic pathway in the early phase of detorsion which triggers inflammatory response [17,6] and the pathway was blocked by amlodipine [18,19] administered immediately on detorsion to prevent Calcium-mediated ROS production [20,21]. Thirdly, there is ROS-burst minutes after reperfusion due to leukocyte recruitment to the site of injury which trigger cell death pathway [9] and vitamin E was administered minutes after detorsion to reduce intratesticular ROS production that can trigger cell death pathways. This treatment strategy therefore provides a practical approach that can be used to prevent TIRI-induced testicular damage in humans.

Febuxostat is a non-purine xanthine oxidase (XO) inhibitor [16,8,22]. It has been reported to reduce ischemia-reperfusion injury via its suppressive effect on xanthine oxidase-reactive

oxygen species (XO-ROS) production. Amlodipine has antioxidant features effective in reducing vascular ischemia-induced damage [21,23] while Vitamin E, is a commonly consumed lipid-soluble antioxidant previously reported to improve spermatogenic function after TIRI induction in rats [24,25]. This study therefore investigates the effect of sequential administration of febuxostat, amlodipine and vitamin E on attenuating oxidative stress induced testicular damage following TIRI.

## 2. MATERIALS AND METHODS

### 2.1 Experimental Animals

Ninety male Wistar rats, weighing 120-150 g were obtained from commercial dealer in Ogbomosho. They were acclimatized for 2 weeks before the commencement of the experiment, and kept throughout the experiment in a well-ventilated plastic cages in the Animal House (temperature 28-31°C; photoperiod:12-h natural light and 12-h dark; humidity:50-55 %) of Faculty of Basic Medical Sciences (FBMS), Ladoke Akintola University of Technology (LAUTECH) with free access to food and water *ad libitum*. The animal handling procedure was done according to the guidelines for the use and care of laboratory animals, as recommended by the animal care and use research ethics committee of LAUTECH, were followed.

### 2.2 Drugs and Reagents

All drugs and reagents used were of high analytical grade. Febuxostat and carboxymethylcellulose (CMC) solution were purchased from TCI chemicals, India (product number: FO840) and LOBA Chemie Pharmaceutical, Ltd. India: Product number: 0253000100. Alpha-Tocopheryl Acetate (Vitamin E) (Gujarat Liquid Pharmacaps Pvt. Ltd, Gujarat, India) and Amlodipine tablets (Swiss Pharmaceutical Pvt. Ltd. Ahmedabad-382445, Gujarat, India).

### 2.3 Experimental Design

Ninety (90) male Wistar rats were divided into 9 groups (n=10) as follows:

**Group 1:** Sham; Rats in this group underwent surgery, without TIRI induction.

**Group 2:** Torsion + Detorsion (TD); Rats underwent left unilateral testicular torsion (TT) for one hour and testicular detorsion followed immediately to induce reperfusion for 56 days.

**Group 3:** Torsion + Febuxostat + Detorsion (TFD); Rats received 5 mg/kg of Febuxostat after 30 minutes of testicular torsion and testicular detorsion followed 30 minutes later, thereafter reperfusion was allowed for 56 days.

**Group 4:** Torsion + Detorsion + Amlodipine (TDA); Rats received 2.5 mg/kg of amlodipine immediately on detorsion that is, one hour after testicular torsion was induced.

**Group 5:** Torsion + Detorsion + Vitamin E (TDV); Rats received 10 mg/kg of vitamin E 30 minutes after detorsion, that is one hour ,thirty minutes after torsion was induced.

**Group 6:** Torsion + Febuxostat + Detorsion+ Amlodipine (TFDA); Rats received 5 mg/kg of Febuxostat after 30 minutes of testicular torsion and 2.5 mg/kg of amlodipine immediately on detorsion.

**Group 7:** Torsion + Febuxostat + Detorsion+ Vitamin E (TFDV); Rats received 5 mg/kg of Febuxostat after 30 minutes of testicular torsion and 10 mg/kg of Vitamin E 30 minutes after detorsion.

**Group 8:** Torsion + Detorsion + Amlodipine + Vitamin E (TDAV); Rats received 2.5 mg/kg of amlodipine immediately on detorsion and 10 mg/kg of vitamin E 30 minutes after detorsion.

**Group 9:** Torsion + Febuxostat + Detorsion + Amlodipine + Vitamin E (TFDAV): Rats received 5 mg/kg of Febuxostat after 30 minutes of testicular torsion and 2.5 mg/kg and 10 mg/kg of Vitamin E immediately on detorsion and 30 minutes after detorsion respectively.

The rats were administered with febuxostat 30 minutes after testicular torsion induction, amlodipine immediately on detorsion (one hour after testicular torsion) and vitamin E 30 minutes after detorsion intraperitoneally once throughout the experiment. Selected doses of febuxostat [16], amlodipine [26] and

vitamin E [27] was based on previous studies.

## 2.4 Experimental Induction of testicular Ischemia-reperfusion Injury

The rats were fasted for 12 hours prior to the experiment. They were weighed and anaesthetized with Ketamine (50 mg/kg) and Xylazine (10 mg/kg) intraperitoneally [28,29]. The rats were restrained on the dissecting board. The left scrotal, perineal and inguinal areas of the rats were shaved and cleaned with methylated spirit. The left testis was firmly grasped and the caudal epididymis was located and used as a reference point. A high left scrotal incision was made to slightly open up the tunica vaginalis to locate the testis. The edges of the tunica vaginalis was clamped with toothed dissecting forceps to produce a tissue plain. The essence of this is to enhance easy returning of the testes back into the scrotum. A gentle pressure was applied to push the left testes out. The gubernaculum testes was located and cut off to free the left testes. The freed left testis was twisted at 720° in a clockwise direction to induce ischemia for one hour. A pouch was created in the scrotum with a long surgical scissors into which an anchoring suture was passed from outside into the inside and attached to the tuft of tissue in-between the testes and epididymis and then passed outward and pulled down to ensure the testes is returned into the scrotum to remain in a twisted state. The incision site was closed up with 2-0 chromic suture. After one hour of torsion, the rats were opened up to untwist the testes to induce reperfusion which lasted for 56 days. This procedure was explained as described by Afolabi et al. [29].

## 2.5 Blood Sample Collection

Fifty-six (56) days after reperfusion, the rats were anaesthetized with ketamine (50 mg/kg). Blood was collected through retro-orbital sinus using heparinized capillary tube and introduced into the plain bottles. The blood collected into the plain bottles were allowed to clot for 15 minutes and then centrifuged at 2500 revolutions per minutes for 15 minutes to obtain serum. The serum was collected into Eppendorf bottles with Pasteur pipettes and refrigerated for further analysis.

## 2.6 Tissue Collection and Preparation of Testicular Homogenate

Testicular tissue were harvested and cleared of adherent tissue. The tissue were cut longitudinally such that one section was fixed inside Bouin's fluid for histological examination of testicular biopsy score and Leydig cell count while other section were homogenized and centrifuged for assay of biochemical parameters.

## 2.7 Biochemical Analysis

Testicular homogenates was used to assess superoxide dismutase (SOD) activity spectrophotometrically using the protocol of Misra and Fridovich, [30]. Catalase activity was assessed spectrophotometrically at 570-610 nm using the method of Sinha, [31], MDA concentration was evaluated according to the method of Adegunlola et al. [32]. Total protein concentration was assessed by the method of Ellam, [33]. Serum nitrite concentration was assessed by checking the nitrite level as described by Fox and Suhre [34] while interleukin-1B was assessed using ELISA kit as described by the manufacturer.

## 2.8 Assessment of Testicular Biopsy Score and Leydig Cell Count

After staining the sections with haematoxylin-eosin stain, they were viewed under light microscope (Omax 40x-2, 500x LED binocular Lab compound Microscope, M82EZ-C50S, China) and the image were observed x 100 magnification. All the cells were counted manually in about 5-10 seminiferous tubules per tissue section or per photomicrograph with the assistance of histology expert. The testicular biopsy score count, an index of complete spermatogenesis was scored using Johnsen's scoring system as previously described by [29]. Each seminiferous tubule evaluated was scored between 1 and 10 depending on the presence or absence of germ cells while large round cells with 1-3 nucleoli in the interstitial space present outside the seminiferous tubules were counted as Leydig cells.

## 2.9 Statistical Analysis

Data were expressed as mean  $\pm$  standard error of mean (Mean  $\pm$  SEM). Analysis of variance with Graph Pad Prism version 7.0 (Graph Pad statistical software, Inc., USA) was used to compare within groups and Tukey's Post-hoc test was used for multiple comparison.  $p < 0.05$  was considered statistically significant.

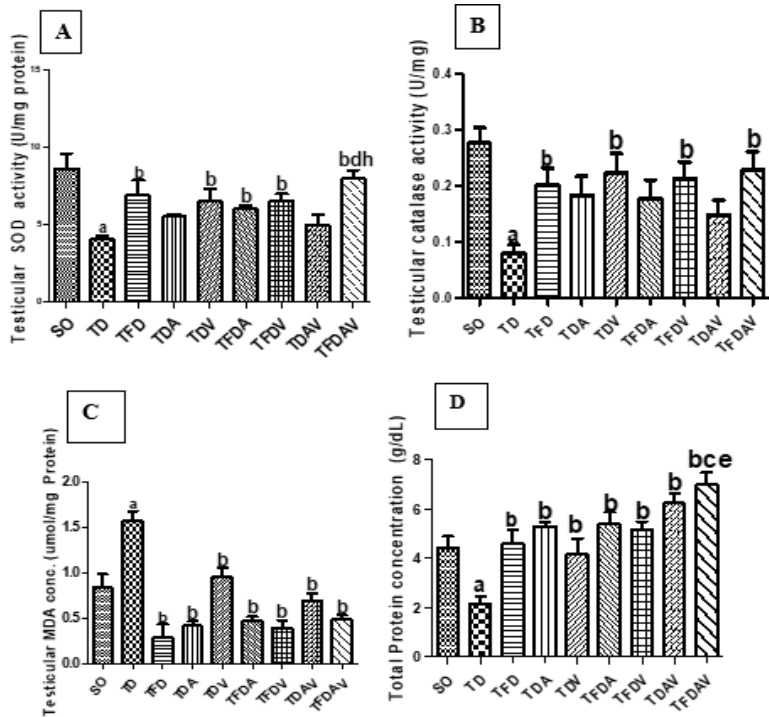
## 3. RESULTS

From the result obtained, the testicular biochemical parameters such as SOD, CAT and total protein were significantly decreased during torsion + Detorsion (Reperfusion injury) while MDA was significantly increased during torsion + Detorsion. Tissue SOD ( $p < 0.01$ ), CAT ( $p < 0.01$ ) and total protein ( $p < 0.01$ ) were significantly decreased in torsion + Detorsion group rats when compared with rats in the sham group while tissue MDA ( $p < 0.01$ ) was significantly increased in torsion + Detorsion group rats when compared to sham group. Tissue SOD was significantly increased ( $p < 0.05$ ) in TFD, TDV, TFDA, TFDV and TFDAV only, CAT was significantly increased in TFD, TDV, TFDV and TFDAV only ( $p < 0.05$ ), total protein was significantly increased in all the treated groups (TFD, TDA, TDV, TFDA, TFDV, TDAV & TFDAV) ( $P < 0.05, 0.01, 0.001$ ) while testicular MDA was significantly decreased in all the groups as well (TFD, TDA, TDV, TFDA, TFDV, TDAV & TFDAV) (febuxostat and vitamin E ( $p < 0.01, p < 0.001$ )) (Fig. 1A, B, C, & D).

IL-1 $\beta$  and serum nitrite were increased ( $p < 0.001$ ) in the Torsion + Detorsion group when compared to sham group but all the treated groups significantly reduced IL-1 $\beta$  and serum nitrite ( $p < 0.05; 0.01; 0.001$ ) (Fig. 2A-B).

Indices of spermatogenesis such as testicular biopsy score and Leydig cell count were reduced in Torsion + Detorsion rats when compared to sham group while all the febuxostat administered group (TFD, TFDA, TFDV & TFDAV) were significantly increased both testicular biopsy score and Leydig cell counts ( $P < 0.01; 0.001$ ) (Fig. 3 A-B).

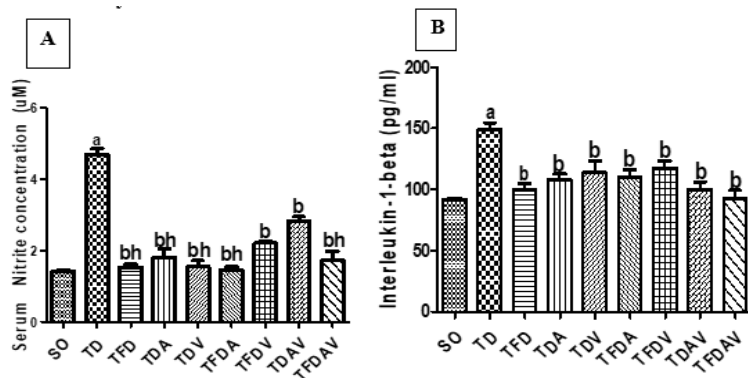
**Biochemical parameters**



**Fig. 1A-D. The effect of sequential administration of febuxostat, amlodipine and vitamin E on superoxide dismutase and catalase activity, malondialdehyde and total protein concentration in male Wistar rats 56 days after TIRI**

*a* represents significance at  $p < 0.01$  when compared to SO.  
*b* represents significance at  $p < 0.05$  when compared to TD.  
*c* represents significance at  $p < 0.01$  when compared to TFD  
*d* represents significance at  $p < 0.001$  when compared to TDA.  
*e* represents significance at  $p < 0.01$  when compared to TDV  
*f* represents significance at  $p < 0.001$  when compared to TFDA.  
*g* represents significance at  $p < 0.01$  when compared to TFDV.  
*h* represents significance at  $p < 0.001$  when compared to TDAV

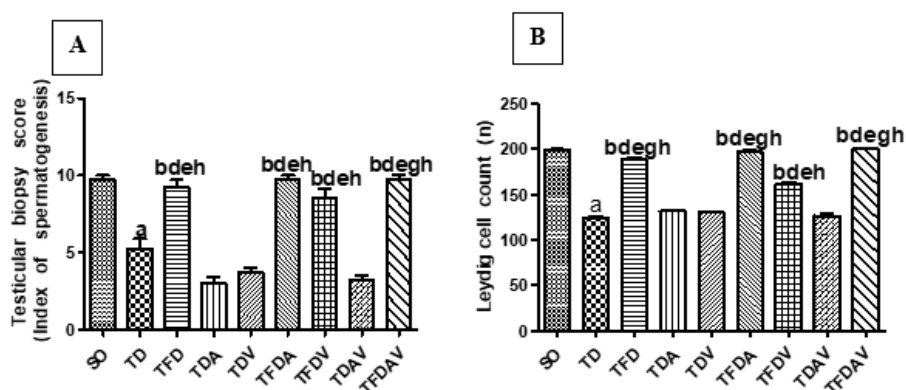
**Inflammatory markers**



**Fig. 2A-B. The effect of sequential administration of febuxostat, amlodipine and vitamin E on serum nitrite concentration and IL-1beta concentration in male Wistar rats 56 days after TIRI**

*a* represents significance at  $p < 0.01$ ;  $0.001$  when compared to SO.  
*b* represents significance at  $p < 0.05$ ;  $0.01$ ;  $0.001$  when compared to TD  
*h* represents significance at  $p < 0.001$  when compared to TDAV

### Testicular biopsy score and Leydig cell counts



**Fig. 3A-B.** The effect of sequential administration of febuxostat, amlodipine and vitamin E on testicular biopsy score and Leydig cell count in male Wistar rats 56 days after TIRI

<sup>a</sup> represents significance at  $p < 0.01$  when compared to SO  
<sup>b</sup> represents significance at  $p < 0.01$  when compared to TD  
<sup>d</sup> represents significance at  $p < 0.001$  when compared to TDA  
<sup>e</sup> represents significance at  $p < 0.001$  when compared to TDV  
<sup>g</sup> represents significance at  $p < 0.05$  when compared to TFDA  
<sup>h</sup> represents significance at  $p < 0.001$  when compared to TDAV

## 4. DISCUSSION

This study shows that long-term testicular ischemia-reperfusion injury (TIRI) after torsion repair can cause infertility in male via impact of oxidative stress and inflammation on spermatogenesis indices. This study also demonstrates the ameliorative effect of sequential administration of febuxostat, amlodipine and vitamin E on the oxidoinflammatory effect of TIRI.

Antioxidants investigated include superoxide dismutase (SOD) and catalase (CAT). In this study, the observed decrease in SOD, and catalase activities in the TD group is an indication of oxidative stress caused as a result of increase depletion of these antioxidants by free radicals generated during reperfusion injury. This is in agreement with the findings [35,36] that reported depletion of antioxidant enzymes following TIRI. These enzymes are crucial for maintaining the redox balance in the body. SOD is an antioxidant enzyme that catalyses the dismutation of superoxide radicals ( $O_2^-$ ) to generate oxygen ( $O_2$ ) and hydrogen peroxides ( $H_2O_2$ ) (Olufunmilayo et al., 2023). The antioxidant activity of catalase is dependent on the degree of SOD activity because it breaks down hydrogen peroxide ( $H_2O_2$ ) into water and oxygen [37].

Administration with only febuxostat or vitamin E, and combination of these treatments as seen in these groups (TFD, TDV, TFDA, TFDV and TFDAV) elevated SOD activity. Febuxostat administered in the ischemic phase of testicular torsion block xanthine-oxidase (XO)-ROS production to preserve SOD activity which is the first line of defense of the testes against free radicals. This is possibly responsible for the elevated activity of SOD in all the febuxostat administered groups (TFD, TFDA, TFDV and TFDAV) in the ischemic phase. Previous studies have also reported the inhibitory effect of febuxostat on XO-ROS production [38,39]. Previous studies have also reported that pretreatment with febuxostat improved antioxidant defense system in diverse organs [22,40].

In addition to this, the vitamin E administered group (TDV) also elevated SOD activity which may be due to its ability to stimulate nuclear factor erythroid 2-related factor 2 (Nrf2) pathway to increase the transcription of SOD and other antioxidant enzymes to protect the testicular cell from excessive ROS generated during TIRI and Vitamin E has also been reported to increase the mobilization of this antioxidant (SOD and others) from the cytosol which is the site of antioxidant production to the mitochondria, the site of ROS generation [41,42]. Through this mechanism, the

febuxostat administered group and the vitamin E administered group was able to restore the depleted SOD activity caused by TIRI.

Catalase activity was also elevated in febuxostat administered groups in the ischemic phase (TFD, TFDV and TFDV) and vitamin E administered (TDV) group only. This result also suggests the effectiveness of febuxostat administered in the ischemic phase to reduce hydrogen peroxide level. Febuxostat administered in the ischemic phase reduce hydrogen peroxide level thereby sparing catalase activity. Previous studies have also reported the suppressive effect of febuxostat against hydrogen-peroxide induced tissue damage [43]. Vitamin E administered (TDV) group also elevated catalase activity due to its ability to inhibit the activity of hydroxyl radicals in the tissue [44].

In this study, the observed reduction in testicular total protein concentration in the TD is also indication of oxidative stress. Proteins are essential biomolecules involved in various cellular processes, including structural integrity, enzymatic catalysis, signalling pathways, and transport mechanisms [45,46]. The depletion of protein level suggests a disruption in protein synthesis, increased protein degradation, or a combination of both, as a consequence of the oxidative stress and cellular damage associated with IRI [44,8].

Ischemia-reperfusion injury induces ER stress, which is characterised by accumulation of misfolded or unfolded proteins in the ER lumen [8]. Prolonged ER stress, as may be the case in this study 56 days after TIRI, can lead to cell death and tissue injury. Also, there can be disruption of cellular energy metabolism, leading to a depletion of ATP levels and subsequent impairment of energy-dependent processes [46]. Moreover, oxidative stress and damage to ribosomes and transcriptional machinery can further compromise protein synthesis [45,44]. Proteins, which are susceptible to oxidation by ROS, can form reversible or irreversible oxidative modifications, such as disulfide bonds, sulfenic acids, and sulfinic/sulfonic acids [47]. These modifications can alter protein structure and function, contributing to cellular dysfunction and tissue injury [47,44]. Sequential administration of febuxostat, amlodipine and vitamin E probably due to their combined antioxidant effect showed better potential in preserving protein levels in the tissue. Febuxostat may reduce oxidative modifications and degradation of proteins [48].

Moreover, it may indirectly promote proper protein folding, contribute to the preservation of protein levels by suppressing ER stress. Amlodipine on the other hand, prevents calcium overload to preserve mitochondrial function and integrity, and thus reduce the production of ROS and subsequent oxidative stress [49]. Vitamin E neutralises ROS generation, thereby preventing the depletion of non-protein thiol and the oxidation of protein thiol, contributing to the maintenance of antioxidant defences and protein integrity.

Malondialdehyde (MDA) is an established marker of oxidative stress in TIRI. This study corroborates this fact as reflected in their significantly higher levels in the TD group. MDA, a marker of lipid peroxidation emanates from the degradation of polyunsaturated fatty acids in cellular membranes [44], Malondialdehyde (MDA) is one of the by-products of lipid peroxidation [6] hence increased MDA levels in the testis homogenate indicated the occurrence of membrane lipid peroxidation and damage via repeated radical chain reactions. Upon reperfusion, XO catalyzes the generation of ROS which can lead to lipid peroxidation and oxidative stress. Administration of febuxostat caused inhibition of XO which attenuate the production of uric acid and consequently ROS generation, thereby mitigating lipid peroxidation and preserving endothelial function [8,48], this may be responsible for the observed reduction in MDA level in the group than TDAV. Amlodipine and vitamin E also help attenuate oxidative stress by scavenging ROS, which in turn preserve the normal levels of MDA [50,51]. Similarly, by maintaining proper calcium levels, amlodipine can preserve mitochondrial function and integrity, reducing the production of ROS and subsequent lipid peroxidation [52].

One of the main features associated with IRI is inflammation, which greatly contributes to tissue damage and compromised function. Serum nitrite levels, which is a storage form of nitric oxide (NO), has been reported to be involved in inflammatory response following TIRI [53]. NO is produced from the oxidation of L-arginine in the vascular endothelium under the catalytic action of endothelial nitric oxide synthase (eNOS) [54]. sNitric oxides (NO) interact with oxygen radicals ( $O_2^-$ ) to form reactive nitrogen species such as peroxynitrites which are more reactive and injurious [55]. Hence, high level of nitrite 56 days after post-reperfusion suggests the occurrence of oxidative damage by reactive nitrogen species



interactions. When TIRI occurs, the inflammatory response includes the release of pro-inflammatory cytokines like interleukin-1 beta (IL-1 $\beta$ ) [6]. During reperfusion, the pro-inflammatory cytokines cause recruitment of neutrophils from blood to the site of injury. This results in the production of reactive oxygen species (ROS), reactive nitrogen species (RNS) and proteolytic enzymes that are highly toxic into the cell and therefore cause tissue damage [56]. In this study, the observed increase in the level of serum NO and IL-1beta suggest heightened inflammatory state resulting from ischemia and subsequent reperfusion injury, and is consistent with previous studies demonstrating their upregulation in ischemia-reperfusion injury (IRI) [53,57].

The decrease in IL-1 $\beta$  level in the groups treated with febuxostat, amlodipine, and vitamin E, either alone or in combination suggest potent anti-inflammatory abilities of these drugs. The anti-inflammatory action of vitamin E can contribute to the attenuation of the inflammatory response associated with TIRI. It has been suggested that xanthine oxidase plays an important role in inflammatory processes and febuxostat can modulate the inflammatory responses by inhibiting ROS production which may contribute to opening of mitochondrial permeability transition pore (mPTP) [6]. Increased mitochondrial calcium level will lead to activation of calcium pyrophosphate which eventually results in increased release of interleukin-1-B [6,58].

Spermatogenesis indices measured include Leydig cell count and testicular biopsy score count. The testicular biopsy score provides a comprehensive assessment for the evaluation of various cells as they are arranged in the seminiferous tubules of the testes [59]. These cells are present in the seminiferous tubules of the testes for the process of sperm production. The observed decrease in Leydig cell count and testicular biopsy score count in TD indicates disruption of differentiation of germinal epithelium (spermatogonia) into spermatozoa, germ cell loss, impaired spermatogenesis and disruption of Leydig cell steroidogenic function attributed to increased ROS production during TIRI which is equally responsible for reduction in testicular biopsy score count observed. Previous studies have demonstrated that testicular ischemia-reperfusion injury results in permanent loss of spermatogenesis [60,61] and ROS generated is the major cause of spermatogenic abnormalities. This finding is consistent with a previous study

[62] that reported TIRI-induced degeneration of germ cells and impaired spermatogenesis.

Elshaari et al. [63] also demonstrated that TIRI-induced ROS was capable of disrupting the steroidogenic function of the Leydig cells. Careful investigation into the treatment groups showed that febuxostat administered in the ischemic phase (TFD, TFDA, TFDV, TFDAV) improved Leydig cell count and testicular biopsy score count 56 days after TIRI. This may be due to suppressive effect of febuxostat on xanthine oxidase-driven ROS production in the ischemic phase of testicular torsion as well as its anti-inflammatory effect against inflammatory responses that raised the intratesticular ROS to triggers germ cell apoptosis. Also, administration of febuxostat in the ischemic phase of testicular torsion may be responsible for the observed improvement in spermatogenesis indices in all the febuxostat administered rats when compared to those that received amlodipine only (TDA), vitamin E only (TDV), amlodipine and vitamin E administered only (TDAV) combined. Since male infertility has been connected to rapidly increased ROS production as reported by Aitken and Baker, [64], the ROS has to be removed from the testicular environment rapidly, it is important to note that maximum protection against ROS is important to sustain Leydig cell and germ cells' integrity [65].

## 5. CONCLUSION

In conclusion, administration of febuxostat 30 minutes after testicular torsion onset, administration of amlodipine immediately on detorsion and administration of vitamin E 30 minutes after detorsion may serve as practicable treatment strategy to the clinical community to prevent TIRI-induced infertility by improving testicular antioxidant enzymes, suppressing inflammation and improving spermatogenesis indices.

## DISCLAIMER (ARTIFICIAL INTELLIGENCE)

I hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

## ETHICAL APPROVAL

This study was approved by animal ethics committee of FBMS, LAUTECH with reference number: ERCFBMSLAUTECH: 033/05/ 2024.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Shimizu S, Martin DT, Dimitriadis F, Keisuke Satoh, Saito S. The effect of ischemic preconditioning and postconditioning on testicular torsion-detorsion injury. *Global Journal of Biochemistry*. 2011a;3:1
2. Eltzschig HK, Eckle T. Ischemia and reperfusion from mechanism to translation. *Nature Medicine*. 2011;17(11):1391-1401.
3. George V, Nick Z. Antioxidants in experimental ischemia-reperfusion injury of the testis: Where are we heading towards? *2017;7(2) 37-45*.
4. Martinon F. Signalling by ROS drives inflammasome activation. *European Journal of Immunology*. 2010;40(3):616-619.
5. Cay A, Alver A, Kucuk M, Isik O, Eminagaoglu MS, Karahan SC. The effects of N-acetylcysteine on antioxidant enzyme activities in experimental testicular torsion. *J Surg Res*. 2006;131:199-203.
6. Kalogeris T, Baines CP, Krenz M, Korthuis RJ. Cell biology of ischemia/reperfusion injury. *International Review of Cell and Molecular Biology*. 2012;298:229-317.
7. Lysiak JJ, Nguyen QA, Kirby JL, Turner TT. Ischemia-reperfusion of the murine testis stimulates the expression of pro-inflammatory cytokines and activation of c-jun N-terminal kinase in a pathway to E-selectin expression. *Biol Reprod*. 2003; 69(1):202-210.
8. Granger DN, Kvietys PR. Reperfusion injury and reactive oxygen species: The evolution of a concept. *Redox Biology*. 2015;6:524-551.
9. Turner TT, Bang HJ, Lysiak JL. The molecular pathology of experimental testicular torsion suggests adjunct therapy to surgical repair. *Journal of American Urological Association*. 2004;172:2574-2578.
10. Antonuccio P, Minutoli L, Romeo C, Nicòtina PA, Bitto A, Arena S, Altavilla D, Zuccarello B, Polito F, Squadrito F. Lipid peroxidation activates mitogen-activated protein kinases in testicular ischemia-reperfusion injury. *J Urol*. 2006;176:1666-1672.
11. Minutoli L, Antonuccio P, Irrera N, Rinaldi M, Bitto A, Marini H, Pizzino G, Romeo C, Pisani A, Santoro G, Puzzolo D, Magno C, Squadrito F, Micali A, Altavilla D. NLRP3 Inflammasome involvement in the organ damage and impaired spermatogenesis induced by testicular ischemia and reperfusion in Mice. *Journal of Pharmacology and Experimental Therapeutics*. 2015;355:370-380 .
12. Bauernfeind FG, Horvath G, Stutz A, Alnemri ES, MacDonald K, Speert D, Fernandes-Alnemri T, Wu J, Monks BG, Fitzgerald KA, Hornung V. Cutting edge: NF- $\kappa$ B activating pattern recognition and cytokine receptors license NLRP3 inflammasome activation by regulating NLRP3 expression. *The Journal of Immunology*. 2009;183(2):787-791.
13. Molteni M, Gemma S, Rossetti C. The role of toll-like receptor 4 in infectious and noninfectious inflammation. *Mediators of Inflammation*. 2016;333-338.
14. Sezai A, Soma M, Nakata KI, Osaka S, Ishii Y, Yaoita H, Hata H, Shiono M. Comparison of febuxostat and allopurinol for hyperuricemia in cardiac surgery patients with chronic kidney disease (NU-FLASH trial for CKD). *Journal of Cardiology*. 2015;66(4):298-303.
15. Fujii K, Kubo A, Miyashita K, Sato M, Hagiwara A, Inoue H, Ryuzaki M, Tamaki M, Hishiki T, Hayakawa N, Kabe Y. Xanthine oxidase inhibitor ameliorates postischemic renal injury in mice by promoting resynthesis of adenine nucleotides. *JCI Insight*. 2019;4(22).
16. Wang S, Li Y, Song X, Wang X, Zhao C, Chen A, Yang P. Febuxostat pretreatment attenuates myocardial ischemia/reperfusion injury via mitochondrial apoptosis. *Journal of Translational Medicine*. 2015;13(1):1-11.
17. Lysiak JJ, Turner SD, Nguyen QA, Singbartl K, Ley K, Turner TT. Essential role of neutrophils in germ cell-specific apoptosis following ischemia-reperfusion injury of the mouse testes. *Biol Reproduction*. 2001;65:718-25.
18. Ferrari R. Major differences among the three classes of calcium antagonists. *European Heart Journal*. 1997;18 (Supplement A):A56-A70.
19. Mason RP. Mechanisms of plaque stabilization for the dihydropyridine calcium

- channel blocker amlodipine: Review of the evidence. *Atherosclerosis*. 2002;165:191-199.
20. Berkels R, Taubert D, Bartels H, Breitenbach T, Klaus W, Roesen R. Amlodipine increases endothelial nitric oxide by dual mechanisms. *Journal of Pharmacology*. 2004;70:39-45.
  21. Dogan C, Halici Z, Topcu A, Cadirci E, Karakus E, Bayir Y, Selli J. Effects of amlodipine on ischaemia/reperfusion injury in the rat testis. *Andrologia*. 2016; 48(4):441-452.
  22. Khan SI, Malhotra RK, Rani N, Sahu AK, Tomar A, Garg S, Nag TC, Ray R, Ojha S, Arya DS, Bhatia J. Febuxostat modulates MAPK/NF-KBp65/TNF-alpha signalling in cardiac ischemia-reperfusion injury. *Oxid Med Cell Longev*; 2017. DOI :8095825.
  23. Javanmardi S, Azizi S, Mohajeri P, Khordadmehr M. The protective effect of orally administered amlodipine against intestinal ischemia-reperfusion injury in rats. *Iranian Journal of Veterinary Surgery*. 2018;13(2):18-25.
  24. Adekeye AO, Akintayo CO, Sanya JO, Enye IA. Testicular torsion following reperfusion injury in rat model: can vitamin E palliate this injury? *Journal of Modern Drug Discovery and Drug Delivery Research*. 2015;1-3.
  25. Zubair M. Effects of dietary Vitamin E on male reproductive system. *Asian Pacific Journal of Reproduction*. 2017;6(4):145-150.
  26. Sun S, Murray CB, Weller D, Folks L, Moser A. Monodisperse FePt nanoparticles and ferromagnetic FePt nanocrystal superlattices. *Science*. 2000; 287(5460):1989-1992.
  27. Corrales F, Corrales M, Schirmer CC. Preventing intraperitoneal adhesions with vitamin E and sodium hyaluronate/carboxymethylcellulose: A comparative study in rats. *Acta Cirurgica Brasileira*. 2008;23:36-41.
  28. Lorenzini F, Tambara Filho R, Gomes RPX, Martino-Andrad AJ, Erdmann TR, Matias JE. Long-term effects of the testicular torsion on the spermatogenesis of the contralateral testis and the preventive value of the twisted testis orchiepididymectomy. *Acta Cirurgica Brasileira*. 2012;27:388-395.
  29. Afolabi OA, Anyogu DC, Hamed MA, Odetayo AF, Adeyemi DH, Akhigbe RE. Glutamine prevents upregulation of NF-kB signaling and caspase 3 activation in ischaemia/reperfusion-induced testicular damage: An animal model. *Biomedicine & Pharmacotherapy*. 2022;150:113056.
  30. Misra HP, Fridovich I. The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *Journal of Biological Chemistry*. 1972;247(10):3170-3175.
  31. Sinha AK. Colorimetric assay of catalase. *Analytical Biochemistry*. 1972;47(2):389-394.
  32. Adegunlola JG, Afolabi OK, Akhigbe RE, Adegunlola GA, Adewumi OM, Oyeyipo IP, Afolabi AO. Lipid peroxidation in brain tissue following administration of low and high doses of arsenite and L-ascorbate in Wistar strain rats. *Toxicology International*. 2012;19(1):47.
  33. Elam JO. Channeling and over packing in carbon dioxide absorbers. *Anesthesiology*. 1959;19:403.
  34. Fox JB, Suhre FB. The determination of nitrite: A critical review; 1985.
  35. Raedschelders K, Ansley DM, Chen DD. The cellular and molecular origin of reactive oxygen species generation during myocardial ischemia and reperfusion. *Pharmacol Ther*. 2012;133:230-255.
  36. Li ZM. Role of antioxidants in preventing testicular ischemia-reperfusion injury: A narrative review. *European Review for Medical and Pharmacological Sciences*. 2022;26:9126-9143.
  37. Hamam H, Demir H, Aydin M, Demir C. Determination of some antioxidant activities (superoxide dismutase, catalase, reduced glutathione) and oxidative stress level (malondialdehyde acid) in cirrhotic liver patients. *Middle Black Sea Journal of Health Science*. 2022;8(4):506-514.
  38. Shafik AN. Febuxostat improves local and remote organ changes induced by intestinal ischemia-reperfusion in rats. *Dig. Dis. Sci*. 2012;58:650-659.
  39. Nomura J, Busso N, Ives A, Tsujimoto S, Tamura M, So A. Febuxostat, an inhibitor of xanthine oxidase suppresses lipopolysaccharide-induced MCP-1 production via MAPK phosphate -1-mediated inactivation of JNK. *Plos One*. 2013;8(9). DOI:10: 1371.
  40. Farag MM, Ahmed SM, Elhadidy WF, Rashad RM. Superior protective effects of

- febuxostat plus alpha-lipoic acid on renal ischemia/reperfusion-induced hepatorenal injury in rats. *Saudi Journal of Kidney Diseases and Transplantation*. 2019;30(6):1364-1374.
41. Ali SS, Ahsan H, Zia MK, Siddiqui T, Khan FH. Understanding oxidants and antioxidants: Classical team with new players. *Journal of Food Biochemistry*. 2020;44(3):e13145.
  42. Fang J, Yin H, Yang Z, Tan M, Wang F, Chen K, Zuo Z, Shu G, Cui H, Ouyang P, Guo H. Vitamin E protects against cadmium-induced sub-chronic liver injury associated with the inhibition of oxidative stress and activation of Nrf2 pathway. *Ecotoxicology and Environmental Safety*. 2021;208:111610.
  43. Higa Y, Hiasa M, Tenshin H, Nakaue E, Tanaka M, Kim S, Nakaue E, Shimizu S, Tanimoto K, Teramachi J, Harada T, Oda A, Oura M, Sogabe K, Hara T, Sumitani R, Maruhashi T, Yamagami H, Endo I, Matsumoto T. The xanthine oxidase inhibitor febuxostat suppresses adipogenesis and activates Nrf2. *Antioxidant*. 2023;12(1):133.
  44. Ayala A, Muñoz MF, Argüelles S. Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. *Oxidative Medicine and Cellular Longevity*; 2014.
  45. Battelli MG, Polito L, Bolognesi A. Xanthine oxidoreductase in atherosclerosis pathogenesis: Not only oxidative stress. *Atherosclerosis*. 2014;237(2):562-567.
  46. Bhattacharyya A, Chattopadhyay R, Mitra S, Crowe SE. Oxidative stress: An essential factor in the pathogenesis of gastrointestinal mucosal diseases. *Physiological Reviews*. 2014;94(2):329-354.
  47. Ghezzi P. Protein glutathionylation in health and disease. *Biochimica et Biophysica Acta (BBA)-General Subjects*. 2013;1830(5):3165-3172.
  48. Kim H, Baek CH, Chang JW, Yang WS, Lee SK. Febuxostat, a novel inhibitor of xanthine oxidase, reduces ER stress through upregulation of SIRT1-AMPK-HO-1/thioredoxin expression. *Clinical and Experimental Nephrology*. 2020;24:205-215.
  49. Park HH, Han MH, Choi H, Lee YJ, Kim JM, Cheong JH, Ryu JI, Lee KY, Koh SH. Mitochondria damaged by oxygen glucose deprivation can be restored through activation of the PI3K/Akt pathway and inhibition of calcium influx by amlodipine camsylate. *Scientific Reports*. 2019;9(1):15717.
  50. Ganafa AA, Walton M, Eatman D, Abukhalaf IK, Bayorh MA. Amlodipine attenuates oxidative stress-induced hypertension. *American Journal of Hypertension*. 2004;17(9):743-748.
  51. Traber MG, Stevens JF. Vitamins C and E: beneficial effects from a mechanistic perspective. *Free Radical Biology and Medicine*. 2011;51(5):1000-1013.
  52. Peng TI, Jou MJ. Oxidative stress caused by mitochondrial calcium overload. *Annals of the New York Academy of Sciences*. 2010;1201(1):183-188.
  53. Reddy MM, Mahipal SV, Subhashini J, Reddy MC, Roy KR, Reddy PR, Reddanna P. Bacterial lipopolysaccharide-induced oxidative stress in the impairment of steroidogenesis and spermatogenesis in rats. *Reprod Toxicol*. 2006;22:493-500.
  54. Fawzy MS, Aisel BT. Association of serum uric acid levels with components of metabolic syndrome: A cross-sectional analysis in a Saudi adult population. *Int. J. Biomed*. 2020;10:457-466.
  55. Piacenza L, Zeida A, Trujillo M, Radi R. The superoxide radical switch in the biology of nitric oxide and peroxynitrite. *Physiol. Rev*. 2022;102(4):1881-1906.
  56. Liu L, Xu TC, Zhao ZA, Zhang NN, Li J, Chen HS. Toll-like receptor 4 signaling in neurons mediates cerebral ischemia/reperfusion injury. *Molecular Neurobiology*. 2023;60(2):864-874.
  57. Chen CB, Liu LS, Zhou J, Wang XP, Han M, Jiao XY, He XS, Yuan XP. Up-regulation of HMGB1 exacerbates renal ischemia-reperfusion injury by stimulating inflammatory and immune responses through the TLR4 signaling pathway in mice. *Cellular Physiology and Biochemistry*. 2017;41(6):2447-2460.
  58. Amirshahrokhi K. Febuxostat attenuates ulcerative colitis by the inhibition of NF-κB, proinflammatory cytokines, and oxidative stress in mice. *International Immunopharmacology*. 2019;76:105884.
  59. Alukal JP, Khera M, Wheeler TM. Testicular biopsy in male infertility evaluation. *Infertility in the Male*. 2009; 215:66-69.
  60. Turner TT, Brown KJ. Spermatic cord torsion: loss of spermatogenesis despite

- return of blood flow. Biol Reprod. 1993; 49401-407.
61. Mestrovic J, Drmic-Hofman I, Pogorelic Z, Vilovic K, Supe-Domic D, Seselja-Perisin A, Capkun V. Beneficial effect of nifedipine on testicular torsion-detorsion injury in rats. Urology. 2014;84:1194-1198.
62. Al-Saleh I, Coskun S, Al-Rouqi R, Al-Rajudi T, Eltabache C, Abduljabbar M, Al-Hassan S. Oxidative stress and DNA damage status in couples undergoing in vitro fertilization treatment. Reproduction and Fertility. 2020;2:117-139.
63. Elshaari FA, Elfagih RI, Sheriff DS, Barassi IF. Oxidative and antioxidative defense system intesticular torsion/detorsion. Indian J Urol. 2011;27(4):479-484.
64. Aitken RJ, Baker MA. Oxidative stress and male reproductive biology. Reproduction, Fertility and Development. 2004;16(5):581-588.
65. Shimizu S, Saito M, Dimitriadis F, Kinoshita Y, Shomori K, Satoh I, Satoh K. Protective effect of ischaemic post-conditioning on ipsilateral and contralateral testes after unilateral testicular ischaemia-reperfusion injury. Int J Androl. 2011b; 34(3):268-75.

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