

Journal of Experimental Agriculture International

Volume 46, Issue 7, Page 868-874, 2024; Article no.JEAI.118543 ISSN: 2457-0591 (Past name: American Journal of Experimental Agriculture, Past ISSN: 2231-0606)

Variation in Leaf Biochemical Compositon of Guava (*Psidium guajava* L) Hybrids

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: https://doi.org/10.9734/jeai/2024/v46i72640

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/118543

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Cite as: Lokesh, G., B. Tanuja Priya, B. Srinivasulu, M. Siva Prasad, Syed Sadarunnisa, M. Jayaprada, and K. Arunodhayam. 2024. "Variation in Leaf Biochemical Compositon of Guava (Psidium Guajava L) Hybrids". Journal of Experimental Agriculture International 46 (7):868-74. https://doi.org/10.9734/jeai/2024/v46i72640.

Lokesh et al.; J. Exp. Agric. Int., vol. 46, no. 7, pp. 868-874, 2024; Article no.JEAI.118543

Short Research Article

Received: 25/04/2024 Accepted: 28/06/2024 Published: 01/07/2024

ABSTRACT

Interplay of several internal development cues leads to floral induction in perennials like guava, mango and litchi. The transition of plants from the vegetative to the reproductive phase depends on the biochemical composition of the leaves. In view of this, the present study was conducted to know the variation in biochemical composition in non-flowering and flowering guava plants. Seven guava hybrids H1 (ARP selection x Lalit), H2 (Lalit x ARP selection), H3 (Allahabad Safeda x ARP selection), H4 (Lalit x Allahabad Safeda), H5 (Allahabad Safeda x Lalit), H6 (Nagpur Seedless x Allahabad Safeda) and OP (Open pollinated progeny of Allahabad Safeda).) were used for estimation of biochemical constituents in blooming and non-blooming plants. Results revealed that higher levels of total sugars, protein, and carbohydrates were observed in the leaves of non-blooming plants compared to blooming plants. However, compared to non-flowered plants, flowering plants had leaves with a higher amount of total phenol.

Keywords: Guava; carbohydrates; proteins; phenols; total sugars.

1. INTRODUCTION

Psidium guajava L. (2n=22, family Myrtaceae) is grown around the world in tropical and subtropical climates. Guava is a member of the Myrtaceae family, which has around 5650 species and 150 genera [1]. Guava is grown over 292 thousand hectares in India, with a production of 4361 thousand metric tonnes [2]. Major guava producing states are Maharashtra, Bihar, Uttar Pradesh, Gujarat, Madhya Pradesh, Odisha, Andhra Pradesh and Punjab. Guava is grown over an area of 9.53 thousand hectares with an annual production of 229.78 thousand metric tonnes in Andhra Pradesh [3]. Vitamin A, thiamin, riboflavin and ascorbic acid content are abundant in edible guava fruits. The leaves of the guava plant are used to cure diabetes, diarrhea, cancer, neurodegenerative, gastrointestinal, and cardiovascular diseases, whereas dried and ripened fruits are considered the best remedy for dysentery [4,5,6,7,8].

Flowering plays an essential role for successful consecutive production of fruit crops. Understanding the role of biochemical constituents like carbohydrates, proteins, total sugars, and total phenols regarding flower initiation is an important aspect during the initial stage of hybrid evaluation. Plants produced phenolic compounds as secondary metabolites through the metabolism of phenylpropanoid in the pentose phosphate and shikimic acid pathways. Higher amount of carbohydrates was consumed during the beginning and subsequent

development of floral organs [9]. Stimulation of carbohydrate catabolism in pear bud might occur due to lower levels of sugars. Based on the importance of different biochemical constituents during flower initiation, a study was conducted to know the role of carbohydrates, proteins, total sugars, and total phenols in the flowering of guava hybrids.

2. MATERIALS AND METHODS

An experiment was carried out on three-year-old hvbrids of guava at the College of Horticulture, Dr. Y. S. R. Horticultural University, Anantharajupeta, Andhra Pradesh. Hybrids include H1 (ARP selection x Lalit), H2 (Lalit x ARP selection), H3 (Allahabad Safeda x ARP selection), H4 (Lalit x Allahabad Safeda), H5 (Allahabad Safeda x Lalit), H6 (Nagpur Seedless x Allahabad Safeda), and OP (Open pollinated progeny of Allahabad Safeda). A randomized block design was used for the current investigation, with three replications for each treatment. To establish the relationship between leaf biochemical constituents and flowering, leaf samples were collected from flowering and nonflowering hybrids for estimation of carbohydrates, protein, total sugars, and total phenols. Total sugars in leaves were determined by Yemm and Willis [10]. Total phenols from guava leaves were extracted by procedure developed by Singleton et al. [11]. Based on Lowrys [12] method, Protein content was estimated. Total carbohydrates were estimated based on the Anthrone method [13]. The collected data was statistically analyzed by using an analysis of variance (ANOVA) test. All these analyses were processed by using OPSTAT.

3. RESULTS AND DISCUSSION

The overall levels of total sugars, total proteins, total carbohydrates and total phenol varied greatly between blooming and non-blooming hybrid guava plants. The content of biochemical components in the leaves of guava hybrid plants is displayed in Table 1. The total sugar content of non-flowering hybrid leaves ranged from 1.38 to 1.79 mg/g and in blooming hybrid plants, it ranged from 0.77 to 1.58 mg/g. In comparison to flowering hybrids higher total sugars were found in leaves of non-flowering hybrids. Within nonflowering hybrids, the highest total sugar content was observed in H2, H6 and the lowest in H4. Regardless of hybrids, flowering plants showed a lower amount of total sugar compared to nonflowering plants. Cho et al. [14] and Moghaddam and Ende [15] mentioned the role of sucrose in relation to flowering. In vegetative tissues, sugars trigger the proliferation of vegetative organs and produce larger and thicker leaves (19). Sugars produced in mango leaves are a potential driving force for phloem-based transport of a flowering promoter (20). Carbohydrate content in the leaves of non-flowering hybrids ranged from 0.57-1.14 mg/g, whereas it ranged from 0.35-0.71 mg/g in flowering plants. Among nonflowering hybrids, maximum carbohydrates were recorded in H1 and a minimum in H2. During flower-bud differentiation process carbohydrate content of leaves, roots and shoot tips might play an essential role [16]. Adequate amount of carbohydrates is utilized for the initiation and development of floral organs in meristematic region of pear [17]. Regarding protein content in leaves, it varied from 1.42-3.86 mg/g where flowering plants registered a range of 1.26-3.22 mg/g. Among non-flowering hybrids, maximum protein content was observed in H1 and H6. As compared to flowering plants, non-flowering plants showed lower amount of leaf protein content. During flower initiation process genes related to autophagy and degradation of protein content are involved [18].

Leaves of flowering hybrids showed a higher content of total phenols compared to nonflowering ones. It ranged from 5.36-7.43 mg/g in non-flowering plants and 6.52-9.49 mg/g in flowering plants. There was no significant difference among hybrids in non-flowering plants. The highest total phenol content in leaves was observed in OP and the lowest in H2. The mobilization of stored food supplies to shoots. which encourages floral bud differentiation and resulting in flowering. The hormone that causes floral initiation and development in litchi is analogized by phenols [19]. In Mango cv Neelum, the highest level of total phenol content was recorded at flower and fruit bud differentiation [20] which provide supporting evidence on the role of phenol in duava flowering. Phenolic content of fruit buds in mango was stable in undifferentiated (or) scar buds while it increased steadily with advancing flower bud differentiation [21-23].



Fig. 1. Difference of leaf total sugars content in non-flowering over flowering in guava hybrids

Lokesh et al.; J. Exp. Agric. Int., vol. 46, no. 7, pp. 868-874, 2024; Article no.JEAI.118543



Fig. 2. Difference of leaf protein content in non-flowering over flowering in guava hybrids



Fig. 3. Difference of total carbohydrates content in non-flowering over flowering in guava hybrids



Fig. 4. Difference of leaf total phenols content in non-flowering over flowering in guava hybrids

Hybrids	Total sugars (%)		Protein content (mg/g)		Total carbohydrates (mg/g)		Total phenols (mg/g)	
	Non flowering	Flowering	Non flowering	Flowering	Non flowering	Flowering	Non flowering	Flowering
H1	1.58	1.19	4.06	3.10	1.14	0.71	6.28	8.50
H2	1.79	1.58	2.03	2.35	0.57	0.35	5.66	6.52
H3	1.57	0.77	3.86	3.22	0.71	0.43	7.37	9.14
H4	1.38	0.78	3.56	3.00	0.79	0.65	5.36	6.82
H5	1.60	1.20	2.60	2.37	0.73	0.55	6.28	8.52
H6	1.79	1.58	1.42	1.26	0.79	0.61	5.70	6.57
OP	1.60	1.19	2.52	2.25	0.82	0.67	7.43	9.49
C.D.	0.04	0.04	0.20	0.76	0.05	0.11	NS	1.78

Table 1. Total sugars, carbohydrates, proteins and total phenols in flowering and non-flowering guava hybrids

4. CONCLUSION

Assessment of biochemical parameters in the leaves of flowering and non-flowering hybrids revealed that flowering plants registered the highest phenol levels, while non-flowering plants registered high total sugars, carbohydrates, and proteins. Phenols as a secondary metabolite play a major role in plant defence mechanisms and, at higher concentrations, favour flower initiation and prevent flower drop by reducing oxidative stress. Thus, understanding the floral mechanism of guava plants and effectively planning crop improvement programmes is aided by knowledge of leaf biochemical composition in flowering and non-flowering hybrids.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declares that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

 Govaerts R, Sobral M, Ashton P, Barrie F, Holst BK, Landrum LL, Matsumoto K. et al. World checklist of Myrtacea. The Board of Trustees of the Royal Botanic Gardens, Kew; 2008.

Available:http://www.kew.org/wcsp

- 2. Anonymous. National horticulture data base. National Horticulture Board. Ministry of Agriculture, Government of India; 2019.
- 3. Anonymous. National horticulture data base. National Horticulture Board. Ministry of Agriculture, Government of India; 2018.
- Jaiswal VS, Amin MN. Guava and jackfruit. In: Hammerschlag, FA, Litz RE (eds) Biotechnology of perennial fruit crops In: Biotechnology in agriculture, 8. CAB Interenational, Wallingford. 1992;421-31.
- Kamath JV, Rahul N, Ashok CKK, Lakshmi SM. *Psidium guajava*: A review. Int J Green Pharm. 2008;2:9-12.
- 6. Polash, Mohammed Arif Sadik, Md Arif Sakil, Shahida Sazia, Md Atikur Rahman, Md Alamgir Hossain. Production time and

nutritional assessment of garden cress (*Lepidium Sativum* L.) Leaves for Ethno-Botanical Uses in Bangladesh. Asian Journal of Advances in Agricultural Research. 2020;12(4):20-27. Available:https://doi.org/10.9734/ajaar/202 0/v12i430089

- Okonkwo CO, Udodia PS. Chemical composition of three different fractions obtained from the leaves of Cassia Occidentalis. Annual Research & Review in Biology. 2020;34(5):1-6. Available:https://doi.org/10.9734/arrb/2019/ v34i530163
- MacFarlane GR. Leaf biochemical parameters in Avicennia marina (Forsk.) Vierh as potential biomarkers of heavy metal stress in estuarine ecosystems. Marine Pollution Bulletin. 2002 Mar 1;44(3):244-56.
- Ito A, Hayama H, Kashimura Y. Possible roles of sugar concentration and its metabolism in the regulation of flower bud formation in Japanese pear (*Pyrus pyrifolia*). Proc. XXVI IHC – Deciduous Fruit and Nut Trees. Ed. A.D. Webster. Acta Hort. 2004;636:ISHS 365-73.
- 10. Yemm EW, Willis AJ. The estimation of carbohydrates in plant extracts by anthrone. Biochemical Journal. 1954;57: 508-514.
- 11. Singleton VL, Joseph A, Rossi. Colourimetry of total phenolics with phossphomolybdic-phosphotungstic acid reagents. American Journal of Enology and Viticulture. 1965;16:144-158.
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with foin-phenol reagent. Journal of Biochemistry. 1951;193:265-275.
- Hedge JE, Hofreiter BT. In: Methods in carbohydrate chemistry, 7(Eds.,) Whistler, R.L. and Be Miller, J.N., Academic Press, New York. 1962;420.
- 14. Cho L, Pasriga R, Yoon J, Jeon J, An G. Roles of sugars in controlling flowering time. J. Plant Biol. 2018;61:121-130.
- 15. Moghaddam MRB, WM. Ende. Sugars, the clock and transition to flowering. Frontiers in Plant Science. 2013;4:1-6.
- Eshghi S, Tafazoli E, Dokhani S, Rahemi M, Emam Y. Changes in carbohydrate contents in shoot tips, leaves and roots of strawberry (*Fragaria ananassa* Duch.) during flower-bud differentiation. Scientia Horticulturae. 2007;113:255–260.

- 17. Peng SA, Iwahori S. Morphological and cytological changes in apical meristem during flower bud differentiation of Japanese pear, *Pyrus pyrifolia.* J. Jap. Soc. Hort. Sci. 1994;63:313–321.
- Fan K, Zhang Q, Liu M, Ma L, Shi Y, Ruan J. Metabolomic and transcriptional analyses reveal the mechanism of C, N allocation from source leaf to flower in tea plant (*Camellia sinensis*. L). Journal of Plant Physiology; 2018. Available:https://doi.org/10.1016/j.j.jplph.20 18.11.007
- 19. Lal N, Marboh ES, Gupta AK, Kumar A, Anal AKD, Nath V. Variation in leaf phenol content during flowering in litchi (*Litchi chinensis* SONN.). Journal of Experimental Biology and Agricultural Sciences. 2019; 7(6):569–573.
- 20. Kumar M, Ponnuswami V, Jeya Kumar P, Saraswathy S. Influence of season

affecting flowering and physiological parameters in mango. Scientific Research and Essays. 2014;9:1-6.

- Patel PB, Rao MM, Srinivasan CN, Basarkar PW, Nalwadi VG. Physiological and biochemical factors associated with fruit bud differentiation in Alphonso mango: V – total free phenols and polyphenol oxidase. Karnataka Journal of Agricultural Science. 1992;54:338-342.
- Gibson SI. Control of plant development and gene expression by sugar signaling. Current Opinion in Plant Biology. 2005; 8:93–102. DOI: 10.1016/j.pbi.2004.11.003
- 23. Davenport TL. Reproductive physiology of mango. Brazilian Journal of Plant Physiology. 2007;19:363–376.
 DOI: 10.1590/S1677-042020 07000400007

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